

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau



AN

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

| | | | |
|---|--|---|--|
| (51) International Patent Classification 5 : C07K 13/00, 15/28, C12N 15/00, C12P 21/02, A61K 37/02 | | A1 | (11) International Publication Number: WO 94/28016 (43) International Publication Date: 8 December 1994 (08.12.94) |
| (21) International Application Number: PCT/US94/05743 | | (81) Designated States: AU, CA, JP, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). | |
| (22) International Filing Date: 20 May 1994 (20.05.94) | | Published <i>With international search report.</i> | |
| (30) Priority Data: 08/065,844 20 May 1993 (20.05.93) US | | | |
| (71) Applicant: THE TRUSTEES OF COLUMBIA UNIVERSITY IN THE CITY OF NEW YORK [US/US]; West 116th Street & Broadway, New York, NY 10027 (US). | | | |
| (72) Inventors: JESSEL, Thomas, M.; 300 West 108th Street, No. 13A, New York, NY 10025 (US). BASLER, Konrad; Gartenstrasse 19, CH-8700 Kushnacht (CH). YAMADA, Toshiya; 1590 Anderson Avenue, No. 17F, Fort Lee, NJ 07024 (US). | | | |
| (74) Agent: WHITE, John, P.; Cooper & Dunham, 30 Rockefeller Plaza, New York, NY 10112 (US). | | | |
| (54) Title: CLONING, EXPRESSION AND USES OF <i>DORSALIN-1</i> | | | |
| (57) Abstract | | | |
| <p>This invention provides an isolated vertebrate nucleic acid molecule which encodes dorsalin-1. This invention also provides a nucleic acid probe capable of specifically hybridizing with a sequence included within the sequence of a nucleic acid molecule encoding a dorsalin-1. The invention also provides a vector and host vector system for the production of a polypeptide having the biological activity of dorsalin-1 which comprises the above-described vector in a suitable host. This invention also provides a purified vertebrate dorsalin-1. This invention provides a method for stimulating neural crest cell differentiation, a method for regenerating nerve cells, a method for promoting bone growth, a method for promoting wound healing and a method for treating neural tumor using purified dorsalin-1. This invention further provides a pharmaceutical composition comprising purified dorsalin-1 and a pharmaceutically acceptable carrier. Finally, this invention provides an antibody capable of binding to dorsalin-1.</p> | | | |

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

| | | | | | |
|----|--------------------------|----|---------------------------------------|----|--------------------------|
| AT | Austria | GB | United Kingdom | MR | Mauritania |
| AU | Australia | GE | Georgia | MW | Malawi |
| BB | Barbados | GN | Guinea | NE | Niger |
| BE | Belgium | GR | Greece | NL | Netherlands |
| BF | Burkina Faso | HU | Hungary | NO | Norway |
| BG | Bulgaria | IE | Ireland | NZ | New Zealand |
| RJ | Benin | IT | Italy | PL | Poland |
| BR | Brazil | JP | Japan | PT | Portugal |
| BY | Belarus | KE | Kenya | RO | Romania |
| CA | Canada | KG | Kyrgyzstan | RU | Russian Federation |
| CF | Central African Republic | KP | Democratic People's Republic of Korea | SD | Sudan |
| CG | Congo | KR | Republic of Korea | SE | Sweden |
| CH | Switzerland | KZ | Kazakhstan | SI | Slovenia |
| CI | Côte d'Ivoire | LI | Liechtenstein | SK | Slovakia |
| CM | Cameroon | LK | Sri Lanka | SN | Senegal |
| CN | China | LU | Luxembourg | TD | Chad |
| CS | Czechoslovakia | LV | Latvia | TG | Togo |
| CZ | Czech Republic | MC | Monaco | TJ | Tajikistan |
| DE | Germany | MD | Republic of Moldova | TT | Trinidad and Tobago |
| DK | Denmark | MG | Madagascar | UA | Ukraine |
| ES | Spain | ML | Mali | US | United States of America |
| FI | Finland | MN | Mongolia | UZ | Uzbekistan |
| FR | France | | | VN | Viet Nam |
| GA | Gabon | | | | |

CLONING, EXPRESSION AND USES OF DORSALIN-1Background of the Invention

5

Throughout this application various publications are referenced by the names of the authors and the year of the publication within parentheses. Full citations for these publications may be found at the end of the specification immediately preceding the claims. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

15

Inductive interactions that define the fate of cells within the neural tube establish the initial pattern of the embryonic vertebrate nervous system. In the spinal cord, the identity of cell types is controlled, in part, by signals from two midline cell groups, the notochord and floor plate which induce neural plate cells to differentiate into floor plate, motor neurons and other ventral neuronal types (van Straaten et al. 1988; Placzek et al. 1990, 1993; Yamada et al. 1991; Hatta et al. 1991). The induction of floor plate cells appears to require a contact-mediated signal (Placzek et al. 1990a, 1993) whereas motor neurons can be induced by diffusible factors (Yamada et al., 1993). Thus, the fate of different ventral cell types may be controlled by distinct signals that derive from the ventral midline of the neural tube.

The specification of dorsal cell fates appears not to require ventral midline signals since the neural tube still gives rise to dorsal cell types such as sensory relay neurons and neural crest cells after elimination of

-2-

the notochord and floor plate (Yamada et al. 1991; Placzek et al. 1991; Ericson et al. 1992). Moreover, dorsal cell types are found at more ventral positions in such embryos (Yamada et al. 1991; Placzek et al. 1991) 5 suggesting that many or all cells in neural tube have acquired dorsal characteristics. The acquisition of a dorsal fate could represent a default pathway in the differentiation of neural plate cells or a response to inductive factors that are distinct from the ventralizing 10 signals that derive from the notochord and floor plate.

To identify signals that might regulate cell differentiation within the neural tube, genes encoding secreted factors that are expressed in a restricted 15 manner along the dorsoventral axis of the neural tube have been searched. In this application, the transforming growth factor β (TGF β) family have been focused since some of its members have been implicated in the control of cell differentiation and patterning in 20 non-neural tissues. In frog embryos, for example, the differentiation and patterning of mesodermal cell types appears to be controlled, in part, by the action of activin-like molecules (Ruiz i Altaba and Melton, 1989; Green and Smith, 1990; Thomsen et al. 1990; Green et al. 25 1992). In addition, the dorsoventral patterning of cell types in *Drosophila* embryos is regulated by the *decapentaplegic* (*dpp*) gene (Ferguson and Anderson, 1992a,b). The *dpp* protein is closely related to a subgroup of vertebrate TGF β -like molecules, the bone 30 morphogenetic proteins (BMPs) (Wozney et al. 1988), several members of which are expressed in restricted regions of the developing embryos (Jones et al. 1991).

In this application, the cloning and functional 35 characterization of the *dorsalin-1* (*dsl-1*) gene, which

-3-

encodes a novel BMP-like member of the TGF- β superfamily are described. *Dsl-1* is expressed selectively by cells in the dorsal region of the neural tube and its expression in ventral regions appears to be inhibited by 5 signals from the notochord. *Dsl-1* promotes the differentiation or migration of neural crest cells and can prevent the differentiation of motor neurons in neural plate explants. The combined actions of *dsl-1* and ventralizing factors from the notochord and floor plate 10 may regulate the identity of neural cell types and their position along the dorsoventral axis of the neural tube.

-5-

This invention provides a method for regenerating nerve cells in a subject comprising administering to the subject an amount of a purified dorsalin-1 effective to regenerate nerve cells.

5

This invention provides a method for promoting bone growth in a subject comprising administering to the subject an amount of a purified dorsalin-1 effective to promote bone growth.

10

This invention provides a method for promoting wound healing in a subject comprising administering to the subject an amount of a purified dorsalin-1 effective to promote wound healing.

15

This invention provides a method for treating neural tumor in a subject comprising administering to the subject an amount of a purified dorsalin-1 effective to inhibit the tumor cell growth.

20

This invention further provides a pharmaceutical composition for stimulating neural crest cell differentiation comprising an amount of a purified dorsalin-1 effective to stimulate neural crest cell differentiation and a pharmaceutically acceptable carrier.

30

This invention provides a pharmaceutical composition for regenerating nerve cells in a subject comprising an amount of a purified dorsalin-1 effective to regenerate nerve cells and a pharmaceutically acceptable carrier.

35

This invention provides a pharmaceutical composition for promoting bone growth in a subject comprising an amount of a purified dorsalin-1 effective to promote bone growth

-6-

and a pharmaceutically acceptable carrier.

This invention provides a pharmaceutical composition for promoting wound healing in a subject comprising an amount 5 of a purified dorsalin-1 effective to promote wound healing and a pharmaceutically acceptable carrier.

This invention provides a pharmaceutical composition for treating neural tumor in a subject comprising an amount 10 of a purified dorsalin-1 effective to inhibit neural tumor cell growth and a pharmaceutically acceptable carrier.

This invention provides an antibody capable of binding to 15 dorsalin-1. This invention also provides an antibody capable of inhibiting the biological activity of dorsalin-1.

-7-

Brief Description of Figures

Figures 1 A-B Nucleotide and Deduced Amino Acid Sequence of Dorsalin-1 (SEQ. ID No. 1.)

5

The numbering of the protein sequence starts with the first methionine of the long open reading frame. The putative signal sequence is typed in bold letters.

10

The RSKR (SEQ. ID No. 17) sequence preceding the proteolytic cleavage site (arrow) is underlined. The site of insertion of the 10 amino acid c-myc epitope is marked with an asterisk. The accession number for dorsalin-1 is L12032.

15

Figure 2 Dorsalin-1 is a Member of the TGF- β Superfamily

20

(A) Alignment of the COOH-terminal amino acid sequences of dorsalin-1 and some representative members of the TGF- β superfamily. Residues that are identical in at least 4 of the 7 proteins are printed in white on a black background. The 7 conserved cysteine residues are marked with an asterisk. Gaps introduced to optimize the alignment are represented by dashes. Known proteolytic cleavage sites in these proteins are marked with an arrow head. Numbers at the right indicate the number of amino acids present in the protein.

25

30

35

(B) Graphical representation of the
SUBSTITUTE SHEET (RULE 26)

-8-

sequence relationship between members of the TGF- β superfamily. This tree representation has been generated using the program pileup of the GCG software package (Devereux et al., 1984). Underneath each branch the percentage amino acid identity is shown with reference to dorsalin-1. This value was calculated using the local homology algorithm of Smith and Waterman (1981) implemented in the program bestfit (GCG software). For both the tree and the amino acid identities only the sequence of the COOH-terminal domain was used, starting with the first of the seven conserved cysteine residues and ending with COOH-terminal residue. For details of other TGF- β family members see Lee (1990), Lyons et al. (1991), Hoffmann, (1991).

Figure 3 Affinity Purification and Functional Activity of Recombinant Dorsalin-1 Protein

(A) Dorsalin-1^{myc} protein was purified from cos-7 cell-conditioned medium using a MAb 9E10 affinity column. An aliquot of the purified protein (CM) was run on a 15% SDS-polyacrylamide gel and stained with Coomassie Blue. The arrow points to the major product running at a molecular weight of ~15 kDa and minor bands at 45, 47 and 60 kDa are also evident. NH₂-terminal sequencing of the 15 kDa band confirmed its identity as processed

-9-

5

dorsalin-1^{myc} protein. Affinity-purified conditioned medium obtained from mock-transfected cos-7 cells did not contain any detectable protein on a Coomassie Blue stained acrylamide gel (not shown). The positions of molecular weight standards (MW) are shown.

10

15

20

25

30

35

(B) Induction of Alkaline Phosphatase Activity in W-20-17 Cells by Dorsalin-1. Conditioned medium was harvested from cos-7 transfected with *dsl-1* cDNA, with the *dsl-1*^{myc} cDNA and added at different dilutions to W20-17 cells for 72h and alkaline phosphatase activity assayed (Thies et al. 1992). As a control for the presence of BMP-like activity in cos-7 cells, medium was also obtained from cells transfected with a c-myc tagged construct encoding the *Drosophila decapentaplegic (dpp)* gene, a related TGF β family member since (see Fig. 2B). Dpp^{myc} is not detectable in the medium of transfected cos-7 cells. Curves are from one of three experiments that produced similar results. Recombinant human BMP-2 (Thies et al. 1992) was used on a positive control in the assay.

Figure 4

Dorsalin-1 mRNA expression in the embryonic chick spinal cord

Panels represent pairs of phase-contrast and dark-field micrographs of sections of embryonic chick neural tube and spinal

-10-

cord, processed for localization of *dorsalin-1* mRNA by *in situ* hybridization with ^{35}S -labelled probe.

5

(A,B) *Dorsalin-1* mRNA is not expressed in neural cells at stages before neural tube closure. The dark field micrograph (B) shows background grain densities.

10

(C,D) *Dorsalin-1* mRNA is expressed at high levels in the dorsal third of the neural tube, beginning at the time of neural tube closure, but not by ventral neural cells or by non-neural cells. This section is taken from a HH stage 10 embryo at the future brachial level.

15

(E,F) The dorsal restriction of *dorsalin-1* mRNA persists in the spinal cord at stages after the onset of neuronal differentiation. Section taken from HH stage 22 embryo, at the brachial level.

20

(G,H) At later stages of spinal cord development (HH St 26) *dorsalin-1* mRNA is restricted to the dorsomedial region of the spinal cord, including but not confined to the roof plate.

25

30

Scale bar: A,B=35 μm , C-F=80 μm , G-H=140 μm .

Figure 5

Regulation of *dorsalin-1* mRNA expression by notochord

-11-

5

(A,B) Phase-contrast and dark-field images of a section of spinal cord from an operated stage 22 embryo but at a level in which there is no grafted tissue. The pattern of *dorsalin-1* mRNA expression is similar to that in unoperated embryos at the same developmental age.

10

(C) Phase-contrast micrograph section from an embryo at the same stage as that shown in A,B, showing the expression of SC1 by motor neurons and floor plate cells, detected by immunoperoxidase histochemistry.

15

20

(D,E) Phase-contrast and dark-field images of a section of spinal cord from an operated stage 22 embryo in which there is a dorsally-located notochord (n). The expression of *dorsalin-1* RNA is suppressed in the presence of a dorsal notochord graft. Similar results were obtained in 2 other embryos.

25

30

(F) Phase-contrast micrograph of an adjacent section to that shown in D,E, showing the ectopic dorsal location of SC1⁺ motor neurons that form a bilaterally symmetric continuous column. SC1⁺ motor axons can be seen leaving the dorsal spinal cord.

35

SC1⁺ floor plate cells are detected at the dorsal midline. The position of the graft d n to chord is indicated (n').

-12-

(G,H) Phase-contrast and dark-field micrographs showing that *dorsalin-1* mRNA expression expands to occupy the entire neural epithelium in embryos from which Hensen's node has been removed at HH stage 10. In this embryo the operation resulted in a splitting of the neural tube and this micrograph has been spliced to restore the ventral apposition of neural tissue. Splitting of the neural tube occurs frequently after removal of Hensen's node (Darnell et al. 1992). A partial or complete ventral expansion of *dsl-1* expression was detected in a total of 5 embryos with Hensen's node removal. A ventral expression of *dsl-1* expression, occupying 60-70% of the spinal cord was also detected after notochord removal in 2 embryos.

20 Scale bar: A-F=90 μ m, G-H=45 μ m.

Figure 6

Induction of Cell Migration from [i]-Neural Plate Explants by Dorsalin-1

25 [i]-Neural plate explants were grown alone or in the presence of *dsl-1*^{myc} (3×10^{-11} M) 48h, and migratory cells analyzed by phase-contrast microscopy and by expression of surface antigens.

30 (A) Phase contrast micrograph of [i]-neural plate explant grown alone for 48h.

35 (B) Phase contrast micrograph of [i]-

-13-

neural plate explant grown in the presence of $dsl-1^{myc}$ for 48h. Many cells have migrated from the explant.

5

(C) Phase contrast micrograph of an [i]-neural plate explant grown in contact with notochord (n) in the presence of $dsl-1^{myc}$ for 48h. Cells still emigrate from the explant although few cells are located in the vicinity of the notochord explant.

10

(D) Expression of HNK-1 by cells induced to migrate from [i]-neural plate explant by $dsl-1^{myc}$.

15

(E) Expression of $\beta 1$ -integrin by cells induced to emigrate from [i]-neural plate explant. About 30% of migratory cells expressed p75, although the levels appeared lower than that detected on neural crest cells derived from the dorsal neural tube.

20

(F) Expression of melanin by cells induced to migrate from quail [i]-neural plate explants by $dsl-1^{myc}$. In these experiments $dsl-1^{myc}$ was removed from after 48h and cultures grown in the presence of chick embryo extract (CEE) for a further 72h. About 10-15% of cells in this bright field micrograph exhibit melanin pigment and typical dendritic morphology. Two different focal planes of the same field are shown to maintain melanocytes in focus. Similar results

25

30

35

-14-

were obtained in 6-8 explants tested. For details see text.

5

(G) Quantitation of cell migration induced by *dsl-1*. [i]np indicates [i]-neural plate explant. nc=notochord, fp=floor plate. Error bars represent the means \pm s.e.m. of migrated cells for 10-26 different explants.

10

Scale bar: A-C=70 μ m, D-F=35 μ m.

15

Figure 7

Induction of *Islet-1* expression in neural plate explants and suppression by *dorsalin-1*

20

(A-C) Normarski (A) and immunofluorescence (B,C) micrographs of stage 9-10 chick [i]-neural plate explant grown for 48h in the absence of notochord or floor plate. *Islet-1*⁺ cells are not detected (B) but there is extensive neuronal differentiation as detected by 3A10 expression (C).

25

30

(D-F) Normarski (D) and immunofluorescence (E,F) micrographs of [i]-neural plate explant grown in contact with stage 26 chick floor plate. Numerous *Islet-1*⁺ cells are present in the [i]-neural plate explant (np), but not in the floor plate explant (fp). The explant also contains many 3A10⁺ cells (F).

35

(G-I) Normarski (G) and immunofluorescence

-15-

5

micrographs (H,I) of [i]-neural plate explant exposed for 48h to floor plate-conditioned medium. Numerous Islet-1⁺ cells (H) and 3A10⁺ neurons (I) are detected.

10

(J-L) Nomarski (J) and immunofluorescence micrograph (K,L) of an [i]-neural plate and floor plate conjugate exposed for 48h to 3×10^{-11} M dorsalin-1^{myc}. No Islet-1⁺ cells are detected (K) whereas the number of 3A10⁺ neurons in the neural plate explant (L) is not obviously different from that in the absence of dorsalin-1^{myc}. In figures D and G, the dashed line outlines the extent of the neural plate (np) explant.

15

20

Scale bar: A-C=70 μ m, D-F=100 μ m, G-I=70 μ m, J-L=100 μ m.

25

30

35

Figure 8

Inhibition of Islet-1⁺ Cells by Dorsalin-1

(A) Histograms showing the induction of Islet-1⁺ cells in [i]-neural plate explants by contact with notochord (nc) or floor plate (fp), and the inhibition of Islet-1⁺ cells by dorsalin-1^{myc} (3×10^{-11} M). Each column represents mean \pm s.e.m. of 10-22 different explants.

(B) Dose-dependent inhibition of Islet-1⁺ cells by dorsalin-1^{myc}. Each point represents mean \pm s.e.m. of 7-23 different explants.

-16-

(C) Induction of Islet-1⁺ cells by floor plate-conditioned medium and the inhibitory action of dorsalin-1^{myc}. Each column represents mean *ts.e.m.* of 7-23 explants.

5

10

15

20

25

30

35

[i]np=[i]-neural plate explant grown alone, +nc=neural plate/notochord conjugate, +fp=neural plate/floor plate conjugate, fpcm=floor plate-conditioned medium.

Figure 9

Potential Functions of Dorsalin-1 in the Control of Cell Differentiation in the Neural Tube

Diagrams summarize the possible mechanisms for establishing the dorsally-restricted expression of dorsalin-1 and potential functions of dorsalin-1 in the regulation of cell differentiation along the dorsoventral axis of the neural tube.

(A) The pattern dorsalin-1 expression appears to be established by early signals from the notochord. (i) Medial neural plate cells respond to signals from the underlying notochord which induce the differentiation of ventral cell types such as floor plate and motor neurons. (ii) Medial neural plate cells are also exposed to signals from the notochord that prevent the subsequent expression of dorsalin-1. The inhibitory signal from the notochord can, in principle, be identical to the

-17-

5 ventralizing signal that induces ventral cell fates. (iii) The medial region of the neural plate gives rise to the ventral neural tube. *Dorsalin-1* expression (shaded area) begins at the time of neural tube closure and is restricted to the dorsal third of the neural tube.

10 (B) *In vitro* assays suggest several possible functions for *dorsalin-1* in the control of neural cell differentiation.

15 (i) *Dorsalin-1* may promote the differentiation of cell types that derive from the dorsal region of the neural tube. *In vitro* studies suggest that neural crest cells represent one population of cells whose differentiation may be influenced by *dorsalin-1*. (ii) The dorsal expression of *dorsalin-1* may define the dorsal third of the neural tube as a domain that is refractory to the long range influence of ventralizing signals from the notochord and floor plate. The ventral boundary of *dorsalin-1* expression suggests that ventral midline-derived signals can influence cells over much of the dorsoventral axis of the neural tube. (iii) *Dorsalin-1* protein may diffuse ventrally to influence the fate of cells in intermediate regions of the neural tube beyond the domain of *dorsalin-1* mRNA expression. Thus, the combined action of *dorsalin-1* and the diffusible ventralizing signal from the notochord and floor plate could specify the fate of cells over the

20

25

30

35

-18-

complete dorsoventral axis of the neural tube.

5

Figure 10 Amino acid comparison of chick dorsalin-1 (B29) and mouse (B29m).

-19-

Detailed Description of the Invention

This invention provides an isolated vertebrate nucleic acid molecule encoding dorsalin-1. As used herein, the 5 term dorsalin-1 encompasses any amino acid sequence, polypeptide or protein having the biological activities provided by dorsalin-1.

In one embodiment of this invention, the isolated nucleic acid molecules described hereinabove are DNA. In a further embodiment, isolated nucleic acid molecules described hereinabove are cDNAs or genomic DNAs. In the preferred embodiment of this invention, the isolated nucleic sequence is cDNA as shown in sequence ID number 10 1. In another embodiment, the isolated nucleic acid molecule is RNA.

This invention also encompasses DNAs and cDNAs which encode amino acid sequences which differ from those of 20 dorsalin-1, but which should not produce phenotypic changes. Alternatively, this invention also encompasses DNAs and cDNAs which hybridize to the DNA and cDNA of the subject invention. Hybridization methods are well-known to those of skill in the art.

25 The DNA molecules of the subject invention also include DNA molecules coding for polypeptide analogs, fragments or derivatives of antigenic polypeptides which differ from naturally-occurring forms in terms of the identity 30 or location of one or more amino acid residues (deletion analogs containing less than all of the residues specified for the protein, substitution analogs wherein one or more residues specified are replaced by other residues and addition analogs where in one or more amino 35 acid residues is added to a terminal or medial position f

-20-

the polypeptides) and which share some or all properties of naturally-occurring forms. These molecules include: the incorporation of codons "preferred" for expression by selected non-mammalian host; the provision of sites for 5 cleavage by restriction endonuclease enzymes; and the provision of additional initial, terminal or intermediate DNA sequences that facilitate construction of readily expressed vectors.

10 The DNA molecules described and claimed herein are useful for the information which they provide concerning the amino acid sequence of the polypeptide and as products for the large scale synthesis of the polypeptide by a variety of recombinant techniques. The molecules are 15 useful for generating new cloning and expression vectors, transformed and transfected prokaryotic and eukaryotic host cells, and new and useful methods for cultured growth of such host cells capable of expression of the polypeptide and related products.

20 Moreover, the isolated nucleic acid molecules are useful for the development of probes to study the neurodevelopment.

25 Dorsalin-1 may be produced by a variety of vertebrates. In an embodiment, a human dorsalin-1 nucleic acid molecule is isolated. In another embodiment, a mouse dorsalin-1 nucleic acid molecule is isolated. In a further embodiment, a chick dorsalin-1 nucleic acid 30 molecule is provided. The plasmid, pKB502, encoding a chick dorsalin-1 was deposited on October 5, 1992 with the American Type Culture Collection (ATCC), 12301 Parklawn Drive, Rockville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the 35 International Recognition of the Deposit of Microorganism

-21-

for the Purposes of Patent Procedure. Plasmid, pKB502 was accorded ATCC Accession number 75321.

Throughout this application, references to specific 5 nucleotides are to nucleotides present on the coding strand of the nucleic acid. The following standard abbreviations are used throughout the specification to indicate specific nucleotides:

10 C=cytosine A=adenosine
 T=thymidine G=guanosine

For the purpose of illustration only, applicants have isolated and characterized dorsalin-1 cDNA clones from chicken and mouse. Similar techniques are applicable to 15 isolate and characterize the dorsalin-1 genes in different vertebrates.

Dorsalin-1 genes may be isolated using the probe generated from the chick dorsalin-1 gene. The mouse and 20 human homologous genes may be cloned by using probe from the chick gene by low stringency screening of the correspondent embryonic spinal cord cDNA libraries. A mouse dorsalin-1 was cloned using the above method. Figure 10 shows a mouse homolog of the dorsalin-1 which 25 reveals extensive conservation at the nucleotide and amino acid level with the chick dorsalin-1. The human dorsalin-1 is likely to be more closely related to the mouse protein than is the chick protein. Thus, it should be straightforward to design oligonucleotide primers to 30 isolate the human dorsalin-1 gene.

This invention provides a nucleic acid molecule comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a 35 sequence included within the sequence of a nucleic acid

-22-

molecule encoding a dorsalin-1. The above molecule can be used as a probe. As used herein, the phrase "specifically hybridizing" means the ability of a nucleic acid molecule to recognize a nucleic acid sequence complementary to its own and to form double-helical segments through hydrogen bonding between complementary base pairs.

10 Nucleic acid probe technology is well known to those skilled in the art who will readily appreciate that such probes may vary greatly in length and may be labeled with a detectable label, such as a radioisotope or fluorescent dye, to facilitate detection of the probe. DNA probe molecules may be produced by insertion of a DNA molecule 15 which encodes dorsalin-1 into suitable vectors, such as plasmids or bacteriophages, followed by transforming into suitable bacterial host cells, replication in the transformed bacterial host cells and harvesting of the DNA probes, using methods well known in the art. 20 Alternatively, probes may be generated chemically from DNA synthesizers.

25 The probes are useful for 'in situ' hybridization or in order to locate tissues which express this gene, or for other hybridization assays for the presence of this gene or its mRNA in various biological tissues.

30 Vectors which comprise the isolated nucleic acid molecule described hereinabove also are provided. Suitable vectors comprise, but are not limited to, a plasmid or a virus. These vectors may be transformed into a suitable host cell to form a host cell vector system for the production of a polypeptide having the biological activity of dorsalin-1.

-23-

This invention further provides an isolated DNA or cDNA molecule described hereinabove wherein the host cell is selected from the group consisting of bacterial cells (such as E.coli), yeast cells, fungal cells, insect cells 5 and animal cells. Suitable animal cells include, but are not limited to Vero cells, HeLa cells, Cos cells, CV1 cells and various primary mammalian cells.

This invention provides a method to identify and purify 10 expressed dorsalin-1. A myc-epitope was introduced into dorsalin-1. This myc carrying dorsalin-1 was linked to an expression vector. Such vector may be used to transfect cell and the distribution of dorsalin-1 in the cell may be detected by reacting myc antibodies known to 15 be reactive to the introduced myc-epitope with the transfected cells which is expressing the dorsalin-1 carrying myc-epitope. Taking advantage of this myc-epitope, dorsalin-1 may be purified by an antibody affinity column which binds with this myc-epitope.

20 In one embodiment, the expression vector, pKB501 (with myc epitope), containing chick dorsalin-1 with a myc-epitope was deposited on October 5, 1992 with the American Type Culture Collection (ATCC), 12301 Parklawn 25 Drive, Rockville, Maryland 20852, U.S.A. under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganism for the Purposes of Patent Procedure. Plasmid, pKB 501 (with myc epitope) was accorded ATCC designation number 75320.

30 The above uses of the myc-epitope for identification and purification of dorsalin-1 should not be considered limiting only to the myc-epitope. Other epitopes with specific antibodies against them which are well known to 35 an ordinary skilled in the art could be similarly used.

-24-

Also provided by this invention is a purified vertebrate dorsalin-1. As used herein, the term "purified vertebrate dorsalin-1" shall mean isolated naturally-occurring dorsalin-1 or protein (purified from nature or 5 manufactured such that the primary, secondary and tertiary conformation, and posttranslational modifications are identical to naturally-occurring material) as well as non-naturally occurring polypeptides having a primary structural conformation (i.e. continuous 10 sequence of amino acid residues). Such polypeptides include derivatives and analogs. In one embodiment, the purified dorsalin-1 is human dorsalin-1.

This invention also provides polypeptides encoded by the 15 above-described isolated vertebrate nucleic acid molecules.

This invention provides a method for stimulating neural crest cell differentiation in a culture comprising 20 administering an amount of the above-described purified dorsalin-1 effective to stimulate neural crest cell differentiation to the culture.

This invention also provides a method for stimulating neural crest cell differentiation in a subject comprising 25 administering to the subject an amount of the above-described purified dorsalin-1 effective to stimulate neural crest cell differentiation.

30 This invention provides a method for regenerating nerve cells in a subject comprising administering to the subject an effective amount of the above-described purified dorsalin-1 effective to regenerate nerve cells.

35 This invention provides a method for promoting bone

-25-

growth in a subject comprising administering to the subject an effective amount of the above-described purified dorsalin-1 effective to promote bone growth.

- 5 This invention provides a method for promoting wound healing in a subject comprising administering to the subject an effective amount of above-described purified dorsalin-1 effective to promote wound healing.
- 10 This invention provides a method for treating neural tumor in a subject comprising administering to the subject an amount of the above-described purified dorsalin-1 effective to inhibit the tumor cell growth. In an embodiment, the neural tumor is neurofibroma. In
- 15 another embodiment, the neural tumor is Schwann cell tumor.

This invention also provides a method for preventing differentiation of motor neurons in a culture comprising administering an amount of purified dorsalin-1 neurons to the culture.

This invention also provides a method for preventing differentiation of motor neurons in a subject comprising administering to the subject an amount of the above-described dorsalin-1 effective to prevent differentiation of motor neurons.

This invention also provides a pharmaceutical composition for stimulating neural crest cell differentiation comprising an amount of purified dorsalin-1 of claim 18 effective to stimulate neural crest cell differentiation and a pharmaceutically acceptable carrier.

35 As used herein, "pharmaceutically acceptable carriers"

-26-

means any of the standard pharmaceutically acceptable carriers. Examples include, but are not limited to, phosphate buffered saline, physiological saline, water and emulsions, such as oil/water emulsions.

5

This invention provides a pharmaceutical composition for regenerating nerve cells in a subject comprising an amount of the above-described purified dorsalin-1 effective to regenerate nerve cells and a pharmaceutically acceptable carrier.

10 This invention provides a pharmaceutical composition for promoting bone growth in a subject comprising an amount of the above-described purified dorsalin-1 effective to promote bone growth and a pharmaceutically acceptable carrier.

15 This invention provides a pharmaceutical composition for promoting wound healing in a subject comprising an amount of the above-described purified dorsalin-1 effective to promote wound healing and a pharmaceutically acceptable carrier.

20 This invention provides a pharmaceutical composition for treating neural tumor in a subject comprising an amount of the above-described purified dorsalin-1 effective to inhibit neural tumor cell growth and a pharmaceutically acceptable carrier. In an embodiment of this pharmaceutical composition, the neural tumor is neurofibroma. In another embodiment of this pharmaceutical composition, the neural tumor is Schwann cell tumor.

25 Also provided by this invention is a method to produce antibody using the above-described purified dorsalin-1.

-27-

Standard procedures for production of antibodies against dorsalin-1 are well-known to an ordinary skilled artisan. A procedure book, entitled "Antibodies, A Laboratory Manual" (1988) by Ed Harlow and David Lane (published by Cold Spring Harbor Laboratory) provides such standard procedures. The content of "Antibodies, A Laboratory Manual" is hereby incorporated in this application.

10 This invention further provides antibody capable of binding to dorsalin-1. In an embodiment, the antibody is monoclonal.

15 This invention further provides an antibody against dorsalin-1 capable of inhibiting the biological activity of dorsalin-1.

20 This invention further provides a method for inhibiting dorsalin-1 activity in a subject comprising administering to the subject an amount of an antibody capable of inhibiting dorsalin-1 activity effective to inhibit the dorsalin-1 activity.

25 This invention also provides a pharmaceutical composition for inhibiting dorsalin-1 activity comprising an amount of antibody capable of inhibiting dorsalin-1 activity effective to inhibit dorsalin-1 activity and a pharmaceutically acceptable carrier.

30 This invention will be better understood from the Experimental Details which follow. However, one skilled in the art will readily appreciate that the specific methods and results discussed are merely illustrative of the invention as described more fully in the claims which

35 follow thereafter.

-28-

EXPERIMENTAL DETAILS

Experimental Procedures

5 RNA Isolation and PCR Amplification

Spinal cord tissue was dissected from 80 embryonic day (E) 2.5 chicks. Poly (A)⁺ RNA (20 μ g) was isolated from this tissue using an oligo (dT)-cellulose spin column (Pharmacia[®]) and 1.5 μ g was used in two first strand cDNA synthesis reactions with either oligo (dT) or random hexanucleotides as primers for the reverse transcriptase reaction. One third of each of the two cDNA reaction mixture was combined and used as template for PCR amplification using 100 pmoles of the following degenerate primers in a reaction volume of 50 μ l:

10 5'TGGAATTCTGG(ACG)A(ACGT)GA(CT)TGGAT(ACT)(AG)T(ACGT)GC
15 3'(SEQ ID No. 10)
and

20 5'GAGGATCCA(AG)(ACGT)GT(CT)TG(ACGT)AC(AGT)AT(ACGT)GC(AG)TG
3'(SEQ ID No. 11)
where degenerate positions are in parenthesis and restriction sites underlined. These oligonucleotides correspond to the dorsalin-1 amino acid positions 339-345
25 and 377-371, respectively. The reaction was cycled twice between 94° (50 seconds), 50° (2 minutes), and 72° (2 minutes), followed by 28 rounds of 94° (50 seconds), 55° (2 minutes), and 72° (1.5 minutes). The reaction products were purified, digested with BamHI and EcoRI, size selected by agarose gel electrophoresis and cloned
30 into the bacteriophage vector M13mp18. 50 clones were picked randomly and analyzed on a sequencing gel by comparing their G ladders. One member of each class was sequenced completely.

-29-

DNA Isolation and Sequencing

An E2.5 chick spinal cord cDNA library of 10^6 independent clones was constructed in lambda ZAPII (Stratagene®) 5 using 5 µg of the poly(A)+ RNA described above. After amplifying the library, 10^6 clones were screened under standard hybridization conditions and a 32 P-labeled PCR probe derived from the 116 bp insert of M13 clone B29 representing the *dorsalin-1* class. Of approximately 25 10 positive clones, 4 were plaque-purified and converted into pBluescript plasmids. Sequence analysis was performed by a combination of primer walking and subcloning of small restriction fragments into M13. The sequence within and adjacent to the long open reading 15 frame was determined on both strands by the dideoxy chain termination method (Sanger et al. 1977) using Sequenase® (U.S. Biochemicals).

DNA Constructs

20

The coding region of *dorsalin-1* was isolated using the two PCR primers ORF-5' (5' TGGAATTCATCGATAACGGAAGCTGAAGC 3'; SEQ ID No. 12) and ORF-3' (5' AGCGTCGACATCGATATTCAAGCATATACTACC 3'; SEQ ID No. 13) and 25 cloned into pBS SK-between the EcoRI and SalI sites. To insert the c-myc epitope (EQKLISEEDL; SEQ. ID No. 18) two internal primers, each encoding half of the c-myc epitope and *dorsalin* sequences from the epitope insertion site (see Figure 1), were used to produce two PCR fragments, 30 one encoding *dorsalin* N-terminal to the insertion site (with primer ORF-5' and the primer 5' GCGAATTCGATATCAGCTTCTGCTCTGCTCCTATGCTTCTCTTGC 3' [SEQ. ID No. 14]) and the other encoding the C-terminal region (w i t h p r i m e r 5' CGGAATTCGATATCGAGGAGGACCTGAACCACTGTCGGAGAACGTC 3'; SEQ 35

-30-

5 ID No. 15 and primer ORF-3'). These two fragments were joined using their primer-derived EcoRV sites and cloned the same way as the unmodified coding region. Using nearby primers this region was sequenced to confirm that no other mutations had been introduced.

10 A truncated coding region was derived from this construct by cleavage with HindIII, blunting the ends with T4 DNA polymerase and subsequent religation. This leads to a frame-shift mutation which replaces the C-terminal 41 residues of dorsalin with 9 unrelated ones. The unmodified, the epitope-tagged and the truncated dorsalin coding regions were then cloned into the Cos-7 cell expression vector pMT21 between the EcoRI and Xhol sites.

15

In Situ Hybridization Histochemistry

20 A *dorsalin-1* cDNA clone was linearized with XbaI (at amino acid position 176) and used to generate a 1 kb [³⁵S]UTP-labeled antisense RNA probe using T7 RNA polymerase. This probe encompasses the 3' part of the cDNA. Chick embryos were fixed in 4% paraformaldehyde and 10 μ m cryostat sections were mounted on 3-aminopropyltriethoxysilane-treated slides. In situ hybridization was performed essentially as described by 25 Wilkinson, et al. (1987) with exposure times ranging from 4 to 10 days. The distribution of *dorsalin-1* mRNA was confirmed by whole-mount in situ hybridization, performed essentially as described by Harland (1991) using a digoxigenin-11-UTP-labeled RNA probe derived from the 30 template mentioned above (not shown).

Chick Embryo Manipulations

35 Notochord grafting and deletion in ovo was performed as

-31-

described by Yamada et al. (1991). For removal of Hensen's node from stage 9-10 chick embryos in ovo, the embryo was visualized by injection of India ink underneath the cavity between the yolk and embryo.

5 Hensen's node was cut out together with underlying endoderm using fine tungsten needles. After the operation, the window was sealed and the embryo was incubated for further 48h at 37°C in the humidified incubator. Embryos were then fixed with 4%

10 paraformaldehyde overnight at 4°C and embedded in paraffin for in situ hybridization as described above.

Cos-7 Cell Transfections

15 Cos-7 cells were transfected by the DEAE-Dextran method as described by Klar, et al. 1992). For small scale cultures 60 to 100 μ m dishes were used and conditioned medium was prepared by incubating cells expressing dorsalin-1 for 48h in 3 or 6 ml of OPTI-MEM (BRL®),

20 respectively. Large-scale transfections for affinity-purification of dorsalin-1 comprised 15 x 150 mm dishes for transfection with *dorsalin*^{myc} DNA (bearing the myc epitope) and an equal number of *dpp* or mock-transfected plates. This yielded 150 ml of *dorsalin*^{myc} conditioned

25 medium and 150 ml of cos-7 conditioned control medium. The BMP-4 expression plasmids was provided by R. Derynck.

Affinity Purification and Sequence Analysis of *dorsalin-1*^{myc}

30 Conditioned medium (50 ml) containing *dsl-1*^{myc} was clarified by centrifugation at 30,000 x g and affinity-purified on 1 ml of a monoclonal 9E10 (anti-myc) antibody column (Affi-Gel, Biorad®). *Dsl-1*^{myc} protein was eluted

35 with 0.1 M glycine-HCl (pH 2.5) and immediately

-32-

neutralized with 3 M Tris base. The eluate was concentrated and desalted over a 2 ml Centricon-10 microconcentrator (Amicon). The protein concentration of the final fraction (volume approximately 130 μ l), as 5 determined by amino acid analysis, was 0.1 μ g/ml.

For SDS-polyacrylamide gel electrophoresis, 10 μ l of concentrated protein was loaded on a 15% Biorad Mini-Protean II gel and stained with Coomassie Blue. 60 μ l 10 was used on a preparative gel and blotted onto Immobilon membrane in the absence of glycine. The blot was stained briefly with Coomassie Blue and the major band at 15 kD was excised and subjected to N-terminal protein 15 sequencing on a Applied Biosystems 470A gas phase sequencer/120A PTH analyzer. The minor protein migrating slightly slower on the gel (at 16.5 kD) was also sequenced and had the identical N-terminus, suggesting that it is an alternately glycosylated form of *dsl-1*. Affinity-purified conditioned medium from mock- 20 transfected cos-7 cells did not contain any detectable protein on a Coomassie-stained acrylamide gel.

The concentration of dorsalin-1^{MYC} used for bioassays was 25 determined on the assumption that all activity resides in the -15 kDa band which represents about 50% of the protein recovered after affinity-purification. The total protein in the affinity-purified fraction determined by amino acid analysis was found to be 100 ng/ μ l, of which 50 ng/ μ l is assumed to represent active protein. The 30 stock concentration of Dsl-1^{MYC} was therefore 3×10^{-6} M. This stock was then diluted 10^5 fold for most assays to give a final condition of 3×10^{-11} M, assuming negligible losses.

Islet-1 Induction Assay

5 The assay for induction of Islet-1⁺ cells was carried out
as described in detail in Yamada et al. 1993. [i]-Neural
plate explants were isolated from Hamburger Hamilton HH
stage 10 chick embryos (Yamada et al. 1993) and grown in
collagen gels alone or with HH stage 10 notochord, HH
stage 26 floor plate or with floor plate-conditioned
10 medium in F12-N3 defined culture medium (Tessier-Lavigne
et al. 1988) at 37°C for 48 to 120h. Floor plate-
conditioned medium was obtained by culturing 30 HH stage
25-26 floor plate fragments in 1 ml of F12 N3 medium for
48h.

15

After incubation, explants were fixed with 4% paraformaldehyde at 4°C for 1-2h, washed with PBS at 4°C and gently peeled from the bottom of the dish and excess collagen gel was trimmed. Explants were incubated with primary antibodies overnight at 4°C with gentle agitation. Rabbit anti-Islet-1 antibodies (Thor et al. 1991, Ericson et al. 1992) and MAb SC1 (Tanaka and Obata, 1984) were used for detection of differentiating motor neurons and MAb 3A10 as a general neuronal marker (Dodd et al., 1988). After washing with PBS for 2h at 22°C, the explants were incubated with Texas Red conjugated goat anti-rabbit antibodies (Molecular Probes) or FITC-conjugated goat anti-mouse Ig (Boehringer Mannheim) for 1-2h. Explants were washed with PBS at 22°C for 2h with at least two changes of buffer and mounted on slides in 50% glycerol with paraphenylenediamine (1 mg/ml). The number of Islet-1⁺ and 3A10⁺ cells was determined on a Zeiss Axiophot microscope equipped with epifluorescence optics. Double labeling with anti-Islet-1 and anti-SCI antibodies was analyzed using Bi Rad confocal microscope.

35

-34-

Analysis of Neural Crest Differentiation

[i]-Neural plate explants from stage 10 chick embryos were grown in collagen gels as described for analysis of Islet-1 induction. The number of migratory cells was determined by phase-contrast microscopy. Cells were scored as migratory if they were greater than two cell body diameters away from the mass of the [i]-neural plate explant. Identification of surface antigens was performed on cultures fixed with 4% paraformaldehyde using MAb 7412 against chick p75 (Tanaka et al. 1989); MAb HNK1 (Abo and Balch, 1981), and MAb JG22 (anti- β 1 integrin; Greve and Gottlieb, 1982). For analysis of melanocyte differentiation, [i]-neural plate explants were isolated from HH st. 10 quail (*Coturnix coturnix japonica*) embryos as described for equivalent chick explants (Yamada et al. 1993) and grown in vitro in collagen gels. Explants were treated with dsl-1^{myc} (3×10^{-11} M) for 48h in F12-N3 medium at which time the medium was removed, explants washed and placed in F12-N3 medium containing 10% chick embryo extract and 10% fetal calf serum for a further 72h. Dsl-1 was removed after 48h because members of the TGF β family have been found to inhibit the differentiation of neural crest cells into melanocytes (Stocker et al., 1991; Roger et al. 1992). CEE and serum were added after 48h to permit the differentiation of neural crest cells into melanocytes (Barofio et al. 1988; Maxwell et al. 1988).

Dorsal neural tube and [i]-neural plate explants grown in dsl-1^{myc} for 48h followed by defined medium lacking CEE or serum for a further 72h gave rise to few, if any, melanocytes. Thus the presence of CEE and serum appears necessary to support melanocyte differentiation under these conditions. When CEE and serum was included in the

-35-

medium from the onset of culture, cells migrated from [i]-neural plate explants and after 120h, melanocytes were observed.

5 To prepare chick embryo extract, white leghorn chicken eggs were incubated for 11 days at 38°C in a humidified atmosphere. Embryos were removed and homogenized in minimal essential medium by passage through a 30 ml syringe, stirred at 40°C for 1h, and then centrifuged for
10 5h at 30,000 x g. The supernatants was collected, filtered and stored at -80°C until used.

Alkaline Phosphatase Induction in W-20-17 Cells

15 Induction of alkaline phosphatase activity by dsl-1 was assayed in W-20-17 cells as described (Thies et al. 1992) using recombinant human BMP-2 as a positive control.

Results

20 Isolation and Characterization of Dorsalin-1

Degenerate oligonucleotides directed against conserved sequences present in the subfamily of TGF- β members that includes the BMPs, Vg1 and dpp were used to isolate novel members of the TGF- β family (Wharton et al., 1991). Oligonucleotides were used as primers in a polymerase chain reaction (PCR) to amplify sequences derived from HH stage 16-18 (embryonic day 2.5) chick spinal cord cDNA.
25 The PCR products were cloned and 37 of 50 clones had inserts encoding Vg-1/dpp/BMP-related peptides. Although most clones encoded chick homologues of previously characterized BMP genes, one class encoded a novel sequence. A 116 bp fragment encoding this sequence was
30 used as probe to screen an E 2.5 chick spinal cord cDNA
35

-36-

library and to define a clone containing a 3.5 kb insert with an open reading frame that encoded a protein of 427 amino acids (Fig. 1).

5 The predicted amino acid sequence identifies this protein, dorsalin-1 (dsl-1), as a new member of the TGF- β superfamily. The N-terminal domain of dsl-1 contains a stretch of hydrophobic residues that could serve as a signal sequence. A comparison of COOH-terminal 109 amino acids with those of other members of this family reveals that dsl-1 contains most of the conserved amino acids present in the other family members, including seven characteristic cysteine residues (Fig. 2A). The structure of TGF- β 2 (Daopin et al., 1992; Schlunegger and Grutter, 1992) suggests that in dsl-1, intrachain disulfide bonds are formed between cysteines 7 and 73, 36 and 106, 40 and 108, and that cysteine 72 is involved in dimer stabilization through formation of an interchain disulfide bond. The NH² terminal domain of the dsl-1 precursor does not exhibit any significant similarity to other members of the TGF- β -family.

20 Dsl-1 is more related to members of the Vg-1/dpp/BMP subfamily than to the TGF- β , activin or MIS subfamilies (Fig. 2B). Given the high degree of sequence conservation of individual members of the BMP family identified in different species (Fig. 2), the divergence in sequence between dsl-1 and mammalian TGF- β family members suggests that the dsl-1 gene encodes a novel member of this superfamily. The sequence of the mouse dsl-1 gene (Cox and Basler, unpublished findings) supports this idea.

25 As with other family members, the conserved COOH-terminal region is immediately preceded by a series of basic

-37-

residues that could serve as a site for proteolytic cleavage of the precursor protein (Celeste et al., 1990; Barr, 1991). An epitope-tagged derivative, dsl-1^{myc}, which contains a 10 amino acid insert derived from the human c-myc proto-oncogene (Evan et al., 1985) was generated to determine the site of cleavage of the dsl-1 precursor. The c-myc sequence was inserted two residues upstream of the first conserved cysteine in a region of the protein that exhibits no conservation with other members of the TGF- β family (Fig. 2A). cDNAs encoding native and epitope-modified dsl-1 were cloned into the expression vector pMT 21 and transfected separately into cos-7 cells.

Medium from cells transfected with the epitope-modified dsl-1 construct was passed over a MAb 9E10 (Evan et al., 1985) anti c-myc affinity column. Affinity purified proteins were analyzed by gel electrophoresis, revealing a major 15 kDa band and minor bands at 45, 47 and ~60 kDa (Fig. 3A). The bands at 45 and 47 kDa correspond in size to those predicted for the unprocessed dsl-1 protein and the 15 kDa band to that expected for a proteolytically-cleaved product. To establish the identity of the 15 kDa band and to determine the site for proteolytic cleavage of the precursor protein, the 15 kDa band was blotted onto Immobilon membranes and subjected to sequence analysis. The NH₂-terminal sequence obtained, SIGAEQKLIS (SEQ ID No. 16), corresponds to residues 319-322 of the predicted dsl-1 sequence followed by the first 6 residues of the human c-myc epitope. This result shows that the R-S-K-R (SEQ ID No. 17) sequence at residues 315-318 is the site of proteolytic processing of the dsl-1 precursor (arrow in Fig. 1), at least in the presence of the c-myc peptide.

-38-

To determine whether recombinant *dsl-1* secreted by cos-7 cells has BMP-like activity, a biochemical assay of osteoblast differentiation was used in which BMPs induce alkaline phosphatase activity (Thies et al. 1992).

5 Recombinant BMP-2 produced a dose-dependent increase in alkaline phosphatase activity in W-20-17 osteoblast cells over a concentration range of 10-1000 ng/ml (not shown; Thies et al. 1992). Conditioned-medium obtained from cos-7 cells transfected with *dsl-1* produced an increase

10 in alkaline phosphatase similar to that of BMP-2 at dilutions of 1:10 to 1:1000 (Fig. 3B). Moreover, medium derived from cos-7 cells transfected with *dsl-1^{MYC}* cDNA, was effective as medium derived from cells transfected with unmodified *dsl-1* cDNA (Fig. 3B). In control

15 experiments, cos-7 cells were transfected with a c-myc tagged version of the *Drosophila decapentaplegic* (*dpp*) gene, which encodes a related TGF- β family member (Fig. 2b). Cos-7 cells do not secret *dpp* protein (Basler, unpublished observations) and medium derived from *dpp* transfectants did not induce alkaline phosphatase

20 activity, providing evidence that cos-7 cells subjected to the same transfection protocol do not secrete a BMP-like activity (Fig. 3B). These results show that *dsl-1* can be expressed in cos-7 cells in functional form, that

25 *dsl-1* mimics the activity of BMPs in this assay and that the activity of *dsl-1* is not reduced by insertion of the c-myc peptide.

Expression of *dsl-1* RNA in the Developing Nervous System

30 *Dsl-1* mRNA was localized in developing chick embryos by *in situ* hybridization to examine the expression of *dsl-1* during neural development. *Dsl-1* mRNA was not expressed by cells in the neural plate (Figs. 4A,B) and first

35 appeared at the time of closure of the neural tube. At

-39-

this stage, *dsl-1* was expressed at high levels in the dorsal third of the neural tube but was absent from more ventral regions (Figs. 4C,D). *Dsl-1* mRNA was restricted to the nervous system at this stage of development (not shown).

The restricted expression of *dsl-1* mRNA in the spinal cord persisted after the onset of neuronal differentiation (Figs. 4E-F), and by E5, the latest stage examined, the domain of expression of *dsl-1* mRNA was confined to the dorsomedial region of the spinal cord including, but not restricted to, the roof plate (Figs. 4G,H). *Dsl-1* mRNA was also expressed in dorsal regions of the hindbrain after neural tube closure (not shown). From E3 to E5, the only non-neural tissue types that expressed detectable levels of *dsl-1* mRNA were kidney and myotomal cells (not shown) although the level of mRNA expression in these tissues was much lower than that in the nervous system.

20

Regulation of *Dsl-1* Expression by the Notochord

The expression of antigenic markers that are restricted to dorsal neural tube cells is regulated by signals from the notochord and floor plate (Yamada et al. 1991; Placzek et al. 1991) raising the possibility that *dsl-1* mRNA expression is controlled in a similar manner. To examine this possibility, segments of stage 10 chick notochord were grafted into the lumen of the neural groove of host embryos prior to the onset of *dsl-1* mRNA expression. Embryos were incubated for a further 48h, during which time the graft was displaced dorsally, such that it is eventually located at the dorsal midline of the neural tube and spinal cord. *Dsl-1* mRNA expression, determined by *in situ* hybridization, was absent from the

-41-

in extreme cases included the entire dorsoventral extent of the neuroepithelium (Figs. 5G,H). In a second series of experiments, the notochord was removed from the caudal region of stage 10 embryos, which were then permitted to 5 develop for an additional 48h. At levels of the spinal cord lacking a floor plate and motor neurons, as assessed by SC1 labelling, the domain *dsl-1* expression expanded ventrally to occupy about two thirds of the spinal cord, although, the most ventral region never expressed *dsl-1* 10 (not shown). The more limited ventral expansion of *dsl-1* observed after removal of the notochord compared with Hensen's node removal is consistent with other studies (Yamada et al. 1993) suggesting that ventralizing signals from the notochord begin to act soon after the neural 15 plate has formed.

Taken together, these experiments suggest that the expression of *dsl-1* mRNA in ventral regions of the neural tube is normally inhibited by signals from the notochord.

20

Dsl-1 Regulates Neural Differentiation In Vitro

The dorsal restriction of *dsl-1* mRNA suggests two ways in which *dsl-1* protein could regulate cell differentiation 25 along the dorso-ventral axis of the neural tube. One function of *dsl-1* could be to promote the differentiation of cell types generated in the dorsal neural tube. A second function of *dsl-1* could be to counteract the influence of ventralizing signals that derive from the 30 notochord and floor plate. The actions of *dsl-1* on the differentiation of defined cell types in neural plate explants grown in vitro have been examined to test the possible functions of *dsl-1*. In the following sections, we provide evidence first that *dsl-1* can promote the 35 differentiation of cells with neural crest-like

-42-

properties and second that *dsl-1* can inhibit the differentiation of motor neurons in response to inductive signals from the notochord and floor plate.

5 **Neural Crest Cell Differentiation:** Neural crest cells are generated from precursors located in the dorsal neural tube (Bronner-Fraser and Fraser, 1988). They can be identified in vitro by their ability to migrate from the neural tube, by their expression of several cell surface markers including the HNK-1 epitope (Maxwell et al. 1988), $\beta 1$ integrin (Delannet and Duband, 1992), the low-affinity neurotrophin receptor subunit p75 (Bernd, 1985; Stemple and Anderson, 1992) and by their ability to differentiate into cell types such as neurons, glial 10 cells and melanocytes (Sieber-Blum and Cohen 1980; 15 Baroffio et al, 1988; Stocker et al. 1991).

To examine whether *dsl-1* might regulate the differentiation or migration of neural crest cells, the intermediate ([i]) region of the neural plate was isolated from stage 10 embryos and grown as explants in vitro (Yamada et al. 1993). As described (Yamada et al. 1993) few cells migrated from [i]-neural plate explants grown in isolation for 48h (Figs. 6A,G). Addition of *dsl-1^{myc}* (3×10^{-11} M) for 48h resulted in a 15-fold increase in the number of cells that migrated from [i]-neural plate explants (Figs. 6B,G). To examine whether these migratory cells share surface properties with chick neural crest cells, cultures grown for 48h in the presence of *dsl-1^{myc}* were labeled with monoclonal 20 antibodies directed against HNK-1, the $\beta 1$ integrin subunit and chick p75. Over 90% of cells that had 25 migrated from the [i]-neural plate explants in the presence of *dsl-1^{myc}* expressed HNK-1 and $\beta 1$ integrin on 30 their surface (Fig. 6D,E) and about 30% expressed p75 35

-43-

(not shown). These results show that cells induced to migrate from [i]-neural plate explants have the properties of neural crest cells.

5 To determine whether the cells that are induced to migrate from [i]-neural plate explants by dsl-1 can differentiate into cell types known to derive from the neural crest, the generation of melanocytes, which can be identified unambiguously in vitro by the presence of
10 melanin pigmentation was studied. In these experiments we used [i]-neural plate explants from quail embryos which exhibit properties in vitro similar to those of equivalently staged [i]-neural plate explants from the non-pigmented chick strain used for all other experiments
15 were used (not shown). Melanocyte differentiation from neural crest cells in vitro has been shown to require permissive factors that can be provided in the form of chick embryo extract (CEE) or serum (Baroffio et al. 1988; Maxwell et al. 1988). [i]-Neural plate explants
20 were therefore grown in dsl-1^{myc} (3×10^{-11} M) for 48h to promote the migration of cells, after which dsl-1^{myc} was removed and the medium supplemented with 10% CEE and 10% fetal calf serum and grown for a further 72h. Under these conditions, 10-15% of the cells that had emigrated
25 from [i]-neural plate explants expressed melanin pigment and exhibited dendritic morphology (Fig. 6F) indicating the presence of melanocytes. Control experiments showed that addition of CEE and serum after exposure of [i]-neural plate explants to dsl-1^{myc} for 48h did not increase further the number of migratory cells (not shown).
30 Moreover, melanocytes were not observed when [i]-neural plate explants were exposed to medium containing CEE and serum for 72h in the absence of dsl-1^{myc} (not shown). These results indicate that cells induced to migrate from
35 [i]-neural plate explants by dsl-1^{myc} can differentiate

-44-

into at least one cell type known to derive from the neural crest.

5 In contrast to neural crest cells that derive from the dorsal neural tube d]-neural plate explants (Yamada et al. 1993), cells that had been induced to migrate from [i]-neural plate explants by dsl-1^{myc} did not express neuronal markers or exhibit neuronal morphology when examined after 48h (not shown). This result suggest that 10 dsl-1 can promote the initial differentiation of neural crest cells from neural plate cells, but that dsl-1 alone does not support the subsequent differentiation of these cells into neurons.

15 The presence of migratory neural crest-like cells was also monitored to address the fate of cells in [i]-neural plate explants that have been exposed both to ventralizing signals and to dsl-1^{myc}. [i]-Neural plate explants grown in contact with the notochord or floor plate for 48h in the presence of dsl-1^{myc} (3×10^{-11} M) exhibited a 12-15 fold increase in the number of migratory cells, similar to that observed when isolated [i]-neural plate explants were exposed to dsl-1^{myc} (Fig. 6G). These cells also expressed HNK-1, 81 integrin and p75 on their surface (not shown). These findings suggest that 20 dsl-1^{myc} promotes the initial differentiation of neural crest cells in the presence of ventralizing signals from the notochord and floor plate.

25 At present, the lack of selective markers has forbidden studies of whether dsl-1 promotes the differentiation of other neural cell types that derive from the dorsal neural tube.

30

35 Regulation f Motor Neuron Differentiation: To examine

-45-

whether *dsl-1* also influences the differentiation of ventral cell types, expression of the LIM homeodomain protein Islet-1 (Karlson et al 1990; Ericson et al. 1992), which provides a marker for the induction of motor 5 neurons in [i]-neural plate explants in response to diffusible signal from the notochord or floor plate was monitored (Yamada et al., 1993).

[i]-Neural plate explants grown in vitro for 48h 10 contained few (usually <5) Islet-1+ cells (Figs. 7A,B;8A,C). In contrast, [i]-neural plate explants grown in contact with notochord or floor plate exhibited a 50-100-fold increase in *Isl-1*+ cells (Figs. 7D,E;8A). Addition of *dsl-1*^{myc} to recombinates of [i]-neural plate 15 with notochord or floor plate produced a concentration-dependent decrease in the number of Islet-1+ cells present in explants (Figs. 7J,K;8A,B). At concentrations of *dsl-1*^{myc} of $3 \times 10^{-11} M$ or greater, the differentiation of Islet-1+ cells was suppressed by over 95% (Fig. 8B). 20 *Dsl-1*^{myc} also abolished the expression of SC1 from regions of the [i]-neural plate explant distant from the junction with the inducing tissue (not shown) suggesting that *dsl-1*^{myc} suppresses motor neuron properties other than *Isl-1*. Addition of *dsl-1*^{myc} to neural plate explants grown alone 25 did not induce Islet-1+ cells (not shown).

A truncated *dsl-1* cDNA in cos-7 cells was expressed and compared its activity with that of native *dsl-1* or *dsl-1*^{myc} to control for the presence of cos-7 cell-derived 30 inhibitory contaminants in preparation of affinity-purified *dsl-1*^{myc}. The induction of Islet-1+ cells by floor plate was suppressed over 95% by a 1:1000 dilution of conditioned medium from cos-7 cells transfected either with unmodified *dsl-1* or with *dsl-1*^{myc} cDNAs (not shown). 35 In contrast, medium derived from cos-7 cells expressing

-46-

the truncated *dsl-1* cDNA did not significantly reduce the number of Islet-1+ cells induced by floor plate (364±62) cells in the absence and 287±45 cell in the presence of medium containing truncated *dsl-1*, mean ±s.e.m., n=4, 5 p>0.10).

Dsl-1 could inhibit the generation of Islet-1+ cells by preventing [i]-neural plate cells from responding to inductive signals or by inhibiting the production of this 10 signal by the notochord and floor plate. The effects of *dsl-1^{myc}* on Islet-1+ cells in [i]-neural plate explants exposed to floor plate-conditioned medium were examined to distinguish these possibilities (Yamada et al. 1993). A 1:10 dilution of floor plate-conditioned medium 15 produced a ~30 fold increase in the number of Islet-1+ cells (Figs. 7G,H;8C). Addition of both *dsl-1^{myc}* and floor plate-conditioned medium to neural plate explants grown alone resulted in a 76% decrease in the number of Islet-1+ cells (Fig. 8C). This result indicates that the 20 inhibition of Islet-1+ cells results, at least in part, from a direct action of *dsl-1* on [i]-neural plate cells.

To examine whether the suppression of Islet-1+ cells is 25 accompanied by a more general inhibition of neuronal differentiation, explants processed for Islet-1 expression were double-labelled with MAb 3A10, a general neuronal marker (Furley et al., 1990). Although the labelling of both cell bodies and axons by 3A10 made it difficult to count the number of neurons accurately, 30 there was no obvious difference in the number of 3A10+ cells in [i]-neural plate explants exposed to concentrations of *dsl-1^{myc}* that almost completely suppressed the differentiation of Islet-1+ cells (Compare Figs. 7I and 7L). These results show that extensive 35 neuronal differentiation still occur under conditions in

-47-

which the induction of Islet-1+ cells is suppressed.

Experimental Discussion

5

Dorsoventral patterning within the neural tube appear to begin at the neural plate stage and to involve the action of both contact-mediated and diffusible inductive signals that derive initially from the notochord and later from 10 the floor plate. A contact-mediated signal appears to be required for floor plate differentiation whereas motor neuron differentiation can be induced by diffusible factors (Placzek et al. 1993; Yamada et al. 1993). The specification of dorsal cell types may, however, require 15 different factors since dorsal cell types persist in the spinal cord of embryos in which the notochord and floor plate have been eliminated.

20

To begin to define factors involved in specifying the fate of cells in the dorsal neural tube, a novel member of the TGFB gene family, *dorsalin-1* (*dsl*), the expression of which is restricted to the dorsal neural tube was cloned and characterized. The dorsal restriction in expression of *dsl-1* appears to be established by signals 25 from the notochord which act on overlying neural plate cells prior to the onset of *dsl-1* transcription to prevent ventral expression of the gene after closure of the neural tube (Fig 9A). The persistence of *dsl-1* mRNA expression in the absence of the notochord and floor plate provides evidence that the expression of genes that 30 are restricted to the dorsal neural tube is independent of ventralizing signals. Dorsal cell fates may be specified by the exposure of neural plate cells to early dorsalizing signals, perhaps from adjacent non-neural 35 ectoderm (Takahashi et al. 1992) which induce the

-48-

potential to express *dsl-1* and other dorsal genes.

Once the dorsal expression of *dsl-1* is established, *dsl-1* protein could function in several different ways to control cell differentiation in the neural tube. First, *dsl-1* may promote the differentiation of cell types that derive from the dorsal neural tube (Fig. 9Bi). Second, the expression of *dsl-1* could ensure that the dorsal neural tube is refractory to ventralizing signals from the notochord (Fig. 9Bii). Finally, *dsl-1* protein could diffuse and influence the fate of cells in more ventral regions of the neural tube (Fig. 9ABiii). The interactions of *dsl-1* and other factors from the dorsal neural tube with ventralizing signals from the ventral midline could, therefore control the identity of cell types and the position at which they are generated along the entire dorsoventral axis of the neural tube.

20 Dsl-1 May Promote Neural Crest Cell Differentiation

One function of *dsl-1* suggested by the pattern of expression of *dsl-1* mRNA could be to promote the differentiation of cell types that are generated in the dorsal neural tube. Neural crest cells constitute one of the major cell types that derive from precursors located in the dorsal neural tube. The present in vitro studies provide evidence that *dsl-1* promotes the initial differentiation of cells with neural crest-like properties from [i]-neural plate explants, but that cells exposed to *dsl-1* alone appear unable to progress to fully differentiated cell types such as neurons or melanocytes. One possible reason for this is that *dsl-1* itself may inhibit neural crest cells from further differentiation. In support of this, TGFB 1 has been shown to inhibit the

-49-

differentiation of neural crest cells into melanocytes (Stocker et al. 1991; Rogers et al. 1992) and to promote the production of extracellular matrix components such as fibronectin (Rogers et al. 1992) that can inhibit 5 neuronal differentiation (Stemple and Anderson, 1992). Alternatively other dorsally-restricted factors that are absent from [i]-neural plate explants may be required for the progression of neural crest cell differentiation.

10 TGFB 1 has also been shown to accelerate the migration of neural crest cells from premigratory regions of the neural tube (Delannet and Duband, 1992). The action of dsl-1 to promote the migration of neural crest-like cells from [i]-neural plate explants differs from this effect 15 in that cells in these explants do not give rise to neural crest cells in the absence of dsl-1 even when maintained in vitro for 96h (Yamada, unpublished observations). Nevertheless, dsl-1 may mimic the ability of TGFB 1 to accelerate neural crest migration and could 20 therefore be involved both in specifying the fate of premigratory neural crest precursors and in inducing the migration of these cells from the dorsal neural tube.

25 It remains unclear whether the differentiation of other classes of dorsal neurons is regulated by dsl-1. Neurons with the properties of dorsal commissural neurons can differentiate in rat neural plate explants grown in isolation (Placzek et al. 1993). Thus it is possible that some dorsal cell types can differentiate 30 independently of dsl-1. Alternatively, neural plate explants grown in vitro may begin to express dsl-1 at levels sufficient to drive the differentiation of some but not all dorsal cell types.

35 Dsl-1 as an Inhibitor of Ventral Cell Type

-50-

Differentiation

Dsl-1 suppresses the differentiation of motor neurons in [1]-neural plate explants exposed to ventralizing signals 5 from the notochord or floor plate. This finding raises the possibility that *dsl-1* interacts with ventralizing signals to control cell fate along the dorsoventral axis of the neural tube. Although, *dsl-1* expression occurs after signals from the notochord and floor plate have 10 begun to specify ventral cell fates (Yamada et al. 1993), its expression precedes the overt differentiation of motor neurons and other ventral neurons (Ericson et al. 1992). Indeed, the first marker of motor neuron 15 differentiation, *Islet-1*, is not expressed until stage 15 (Ericson et al. 1992), or about 18-20h after neural tube closure and the onset of *dsl-1* expression. Thus, in the period between the initial specification and overt differentiation of neurons, *dsl-1* may accumulate to 20 levels that are sufficient to influence neuronal differentiation.

The ability of *dsl-1* to inhibit motor neuron differentiation could be involved in preventing the 25 generation of motor neurons and other ventral cell types in the dorsal neural tube. This presupposes that ventralizing signals from the notochord and floor plate can influence dorsal regions of the neural tube. Secreted factors from the floor plate have been shown to diffuse over long distances through the neuroepithelium 30 (Placzek et al. 1990). Moreover the position of the ventral boundary of the domain of *dsl-1* expression suggests that signals from the notochord can influence at least two third of the neural tube. Thus, expression of *dsl-1* within the dorsal third of the neural tube could 35 make cells in this region refractory to long range

-51-

ventralizing signals from the notochord and floor plate.

The potential contributions of *dsl-1* to cell differentiation along the dorso-ventral axis of the neural tube will also depend on the range of action of *dsl-1* itself. Since *dsl-1* is readily secreted from cells in vitro, *dsl-1* may diffuse ventrally, beyond the domain of *dsl-1* mRNA expression, to influence the response of cells in intermediate regions of the neural tube. Again, the ability of *dsl-1* to antagonize the response of neural cells to ventralizing signals from the notochord and floor plate could be relevant both to the differentiation of motor neurons and to other ventral cell types.

15 Prevention of *Dsl-1* Expression Ventrally May be Required for Ventral Cell Type Differentiation

Dsl-1 promotes neural crest cell migration and inhibits motor neuron differentiation in the presence of the notochord or floor plate. These findings suggest that the actions of *dsl-1* dominate over ventralizing signals. Thus, the inhibition of *dsl-1* expression from ventral regions of the neural tube that is achieved by early signals from the notochord may be necessary for the differentiation of ventral cell types. The absence of ventral cell types in the neural tube of embryos lacking a notochord could, therefore, result either from a ventral expansion in the domain of *dsl-1* expression or from the loss of ventralizing signals. However, in such operated embryos the neural tube is reduced in size (van Straaten and Hekking, 1991), thus, the death (Homma and Oppenheim, 1992) or arrested division (Placzek et al. 1993) of ventral cells could also contribute to the presence of dorsal cell types in regions of the neural tube that appear to be ventral.

-52-

Dsl-1 and the TGFB Family

In addition to *dsl-1*, several other members of the BMP (DVR) subfamily of TGFB-like genes are expressed in the 5 embryonic nervous system. Other BMP-like proteins may therefore mimic the actions of *dsl-1* on neural cell differentiation. In preliminary studies, the induction of motor neurons was found to be also suppressed by cos-7 cell-derived BMP-4 (Basler et al. unpublished). In the 10 spinal cord and hindbrain, the BMP-4 (DVR-4) gene is expressed selectively by cells in the roof plate whereas in the diencephalon, the gene is found at the ventral midline (Jones et al., 1991). The expression of BMP-4 in the ventral diencephalon coincides with, and could 15 perhaps contribute to the absence of motor neurons from the embryonic forebrain. The embryonic distribution of most other BMP genes is not known although Vgr-1 (BMP-6/DVR-6) is expressed by cells immediately adjacent to the floor plate in the spinal cord (Jones et al., 1991) 20 and GDF-1 appears to be expressed widely throughout the embryonic nervous system (Lee, 1990, 1991). Studies to determine whether widely distributed proteins such as GDF-1 mimic the actions of *dsl-1* will be important in evaluating the role of this gene family in neural 25 patterning.

The involvement of *dsl-1* in the control of cell differentiation along the dorsoventral axis of the neural tube extends the range of activities described for 30 members of the TGFB family during embryonic development. Studies in *Xenopus* embryos have provided evidence that activin can control the identity of mesodermal cell types in a concentration-dependent manner (Ruiz i Altaba and Melton, 1989; Green et al. 1992). In addition, the 35 pattern of expression and possible functions of *dsl-1* in

-53-

the neural tube has parallels with that of the decapentaplegic gene (*dpp*) in *Drosophila* embryonic development (Ferguson and Anderson, 1992a,b). Dorsoventral patterning in the early *Drosophila* embryo 5 involves a dorsal restriction of *dpp* expression (St. Johnston and Gelbart, 1987) that is achieved by ventral-midline derived signals that inhibit *dpp* expression ventrally (Ray et al. 1991). Genetic inactivation of this ventral signalling pathway or introduction of *dpp* 10 activity ventrally, changes the fate of cells along the dorsoventral axis of the embryo (Ferguson and Anderson, 1992b). In the neural tube, the dorsal restriction of *dsl-1* mRNA by early signals from the notochord could generate a gradient of *dsl-1* activity along the 15 dorsoventral axis of the neural tube. Alone, or in conjunction with ventralizing signals from the notochord and floor plate, a gradient of *dsl-1* could influence the fate of cells according to their dorsoventral position within the neural tube.

-54-

REFERENCES

5 Abo, T. and Balch, C.M. (1981) A differentiation antigen of human NK and K cells identified by a monoclonal antibody (HNK-1) J.Immun. 127:1024-1029.

10 Barr, P.J. (1991) Mammalian subtilisins: The long-sought after dibasic processing endoproteases. Cell 66:1-3.

15 Baroffio, A., Dupin, E. and LeDouarin, N.M. (1988) Clone-forming ability and differentiation potential of migratory neural crest cells. Proc. Natl. Acad. Sci. 85:5325-5329.

20 Bernd, P. (1985) Appearance of nerve growth factor receptors on cultured neural crest cell. Devl. Biol. 112:145-156.

25 Bronner, Fraser, M. and Fraser, S. (1988) Cell lineage analysis shows multipotentiality of some avian neural crest cells. Nature 335:161-164.

30 Celeste, A.J., Iannazzi, J.A., Taylor, R.C., Hewick, R.M., Rosen, V., Wang, E.A. and Wozney, J.M. (1990) Identification of transforming growth factor B family members present in bone-inductive protein purified from bovine bone. Proc. Natl. Acad. Sci. 87:9843-9847.

35 Daopin, S., Piez, K.A., Ogawa, Y. and Davies, D.R. (1992) Crystal structure of transforming growth factor- β 2: An unusual fold for the superfamily. Science 257:369-373.

Darnell, D.K., Schoenwold, G.C. and Ordahl, C.P. (1992) Changes in dorsoventral but not rostrocaudal regionalization of the chick neural tube in the absence

-55-

of cranial notochord, as revealed by expression of engrailed-2. *Dev. Dyn.* 193:389-396.

5 Delannet, M. and Duband, J-L. (1992) Transforming growth factor- β control of cell-substratum adhesion during avian neural crest cell emigration in vitro. *Dev.* 116:275-287.

10 Devereux, J., Haeberli, P. and Smithies, O. (1984) A comprehensive set of sequence analysis programs for the VAX. *Nucl. Acid Res* 12:387-395.

15 Dodd, J., Morton, S.B., Karagoegeos, D., Yamamoto, M., and Jessell, T.M. (1988) Spatial regulation of axonal glycoprotein expression on subsets of embryonic spinal neurons. *Neuron* 1:105-116.

20 Ericson, J., Thor, S., Edlund, T., Jessell, T.M. and Yamada, T. (1992) Early stages of motor neuron differentiation revealed by expression of homeobox gene Islet-1. *Science* 256:1555-1560.

25 Evan, G.I., Lewis, G.K., Ramsay, G. and Bishop, J.M. (1985) Isolation of monoclonal antibodies specific for human c-myc protooncogene product. *Mol. Cell Biol.* 5:3610-3616.

30 Ferguson, E.L. and Anderson, K.V. (1992a) Localized enhancement and repression of the activity of the TGF- β family member, decapentaplegic, is necessary for dorsal-ventral pattern formation in the *Drosophila* embryo. *Development* 114:583,597.

35 Ferguson, E.L. and Anderson, K.V. (1992b) Decapentaplegic acts as a morphogen to organize dorsal-ventral pattern in the *Drosophila* embryo. *Cell* 71:451-461.

-56-

Furl y, A.J., Morton, S.B., Manalo, D., Karagogeos, D., Dodd, J. and Jessell, T.M. (1990) The axonal glycoprotein TAG-1 is an immunoglobulin superfamily member with neurite outgrowth-promoting activity. *Cell* 5 61:157-170.

Grabowski, C.T. (1956) The effects of the excision of Hensen's Node on the early development of the chick embryo. *J.Exp.Zool.* 133:301-343.

10 Green, J.B. and Smith, J.C. (1990) Graded changes in dose of a *Xenopus* activin A homologue elicit stepwise transitions in embryonic cell fate. *Nature* 347:391-394.

15 Green, J.B., New, H.V. and Smith, J.C. (1992) Responses of embryonic *xenopus* cells to activin and FGF are separated by multiple dose thresholds and correspond to distinct axes of the mesoderm. *Cell* 71:731-739.

20 Greve, J.M. and Gottlieb, D.I. (1982) Monoclonal antibodies which alter the morphology of cultured chick myogenic cells. *J.Cell. Biochem.* 18:221-229.

25 Hamburger, V. and Hamilton, H. (1951) A series of normal stages in the development of chick embryo. *J.Morph.* 88:49-92.

30 Harland, R.M. (1991) In situ hybridization: an improved whole mount method for *Xenopus* embryos. *Meth. Enzymol.* 36:675-685.

35 Hatta, K., Kimmel, C.B., Ho, R.K. and Walker, C. (1991) The cyclops mutation blocks specification of the floor plate of the zebrafish central nervous system. *Nature* 350:339-341.

-57-

Hirano, S., Fuse, S. and Sohal, G.S. (1991) The effect of the floor plate of pattern and polarity in the developing central nervous system. *Science* 251:310-313.

5 Hoffman, F.M. (1991) Transforming growth factor- β -related genes in *Drosophila* and vertebrate development. *Current Opinion in Cell Biology* 3:947-952.

10 Homma, S. and Oppenheim, R.W. (1992) Notochord dependent cell survival in the ventral half of the chick neural tube. *Soc. Neurosci. Abst.* 43.

15 Jones, C.M., Lyons, K.M. and Hogan, B.L.M. (1991) Involvement of bone morphogenetic protein-4 (BMP-4) and Vgr-1 in morphogenesis and neurogenesis in the mouse. *Development* 111:531-542.

20 Karlsson, O., Thor, S., Norbert, T., Ohlsson, H., and Edlund, T. (1990) Insulin gene enhancer binding protein Isl-1 is a member of a novel class of proteins containing both a homeo- and a Cys-His domain. *Nature* 344:879-882.

25 Klar, A., Baldassare, M. and Jessell, T.M. (1992) F-spondin: A gene expressed at high levels in the floor plate encodes a secreted protein that promotes neural cell adhesion and neurite extension. *Cell* 69:95-110.

30 Lee, S.J. (1990) Identification of a novel member (GDF-1) of the transforming growth factor- β superfamily. *Molecular Endocrinology* 4:1034-1040.

Lee, S.J. (1991) Expression of growth/differentiation factor 1 in the nervous system: Conservation of a bicistronic structure. *Proc. Natl. Acad. Sci.* 88:4250-4254.

-58-

Lyons, K.M., Jones, C.M. and Hogan, B.L.M. (1991) The DVR gene family in embryonic development. *Trends Genet.* 7:408-412.

5 Maxwell, G.D., Forbes, M.E. and Christie, D.S. (1988) Analysis of the development of cellular subsets present in the neural crest using cell sorting and cell culture. *Neuron* 1:557-568.

10 Placzek, M., Tessier-Lavigne, M., Yamada, T., Jessell, T.M. and Dodd, J. (1990a). Mesodermal control of neural cell identity: floor plate induction by the notochord. *Science* 250:985-988.

15 Placzek, M., Tessier-Lavigne, M., Jessell, T.M. and Dodd, J. (1990b) Orientation of commissural axons in vitro in response to a floor plate-derived chemoattractant. *Development* 110:19-30.

20 Placzek, M., Yamada, T., Tessier-Lavigne, M., Jessell, T.M. and Dodd, J. (1991) Control of dorso-ventral pattern in vertebrate neural development: induction and polarizing properties of the floor plate. *Development Suppl.* 2:105-122.

25 Placzek, M., Jessell, T.M. and Dodd, J. (1993) Induction of floor plate differentiation by contact-dependent, homeogenetic signals. *Development* 117 (In Press).

30 Ray, R.P., Arora, K., Nusslein-Volhard, C. and Gelbart, W.M. (1991) The control of cell fate along the dorsal-ventral axis of the *Drosophila* embryo. *Development* 113:35-54.

35 Rogers, S.L., Gegick, P.J., Alexander, S.M. and McGuire,

-59-

P.G. (1992) Transforming growth factor- β alters differentiation in cultures of avian neural crest-derived cells: effects on cell morphology, proliferation, fibronectin expression and melanogenesis. Dev. Biol. 5 151:192-203.

Ruizi Altaba, A. and Melton, D.A. (1989) Interaction between peptide growth factors and homeobox genes in the establishment of antero-posterior polarity in frog embryos. Nature 10 341:33-38.

Sanger, F., Nicklen, S. and Coulson, A.R. (1977). DNA sequencing with chain-terminating inhibitors. Proc.Natl.Acad.Sci.USA 15 74:5463-5467.

Schlunegger, M.P. and Grutter, M.G. (1992) An unusual feature revealed by the crystal structure of 2.2A resolution of human transforming growth factor- β 2. Nature 20 358:430.

Sieber-Blum, M. and Cohen, A.M. (1980) Clonal analysis of quail neural crest cells: They are pluripotent and differentiate in vitro in the absence of non-crest cells Devl. Biol. 25 80:96-106.

Smith, T.F. and Waterman, M.S. (1981) Identification of common molecular subsequences. J. of Mol. Biol. 25 147:195-197.

Smith, J.L. and Schoenwolf, G.C. (1989) Notochordal induction of cell wedging in the chick neural plate and its role in neural tube formation. J. Exp. Zool. 30 250:49-62.

Smith-Thomas, L.C. and Fawcett, J.W. (1989) Expression of 35

-60-

Schwann cell markers by mammalian neural crest cells.
Development 105:251-262.

5 Stemple, D.L. and Anderson, D.J. (1992) Isolation of a
Stem Cell For Neurons and Glia from the Mammalian Neural
Crest. Cell 71:973-985.

10 St. Johnson, R.D. and Gelbart, W.M. (1987)
Decapentaplegic transcripts are localized along the
dorsal-ventral axis of the *Drosophila* embryo. The EMBO
Journal 6:2785-2791.

15 Stocker, K.M., Sherman, L., Rees, S. and Ciment, G.
(1991) Basic FGF and TGF- β 1 influence commitment to
melanogenesis in neural crest-derived cells of avian
embryos. Dev. 111:635-645.

20 Takahashi, Y., Monsoro-Burq, A-H., Bontoux, M. and Le
Douarin, N.M. (1992) A role for Quox-8 in the
establishment of the dorsoventral pattern during
vertebrate development. Proc. Natl. Acad. Sci. 89:10237-
10241.

25 Tanaka, H. and Obata, K. (1984) Developmental changes in
unique cell surface antigens of chick embryo spinal motor
neurons and ganglion cells. Devl. Biol. 106:26-37.

30 Tanaka, H., Agata, A. and Obata, K. (1984) A new membrane
antigen revealed by monoclonal antibodies is associated
with motoneuron axonal pathways. Dev. Biol. 132:419-435.

35 Tessier-Lavigne, M., Placzek, M., Lumsden, A.G.S., Dodd,
J. and Jessell, T.M. (1988) Chemotropic guidance of
developing axons in the mammalian central nervous system.
Nature 336:775-778.

-61-

Thies, R.S., Bauduy, M., Ashton, B.A., Kurtzberg, L., Wozney, J.M. and Rosen, V. (1992) Recombinant human bone morphogenetic protein-1 induces osteoblastic differentiation in W-20-17 stromal cells. *Endocrinology* 130:1318-1324.

Thomsen, G., Woolf, T., Whitman, M., Sokol, S., Vaughan, J., Vale, W. and Melton, D.A. (1990) Activins are expressed early in *Xenopus* embryogenesis and can induce axial mesoderm and anterior structures. *Cell* 63:485-493.

Thor, S., Ericson, J., Brannstrom, T. and Edlund, T. (1991) The homeodomain LIM protein *Isl-1* is expressed in subsets of neurons and endocrine cells in the adult rat. *Neuron* 7:881-889.

van Straaten, H.M.W., Hekking, J.W.M., Wiertz-Hoessels, E.L., Thors, F. and Drukker, J. (1988) Effect of the notochord on the differentiation of a floor plate area in the neural tube of the chick embryo. *Anat. Embryol.* 177:317-324.

van Straaten, H.M.W. and Hekking, J.W.M. (1991) Development of floor plate, neurons and axonal outgrowth pattern in the early spinal cord of the notochord-deficient chick embryo. *Anat. Embryol.* 184:55-63.

Wharton, K.A., Thomsen, G.H. and Gelbart, W.M. (1991) *Drosophila* 60A gene, another transforming growth factor β family member, is closely related to human bone morphogenetic proteins. *Proc. Natl. Acad. Sci.* 88:9214-9218.

Wilkinson, D.G., Bailes, J.A. and McMahon, A.P. (1987) Expression of the proto-oncogene *int-1* is restricted to

-62-

specific neural cells in the developing mouse embryo.
Cell 58:79-88.

5 Wozney, J.M., Rosen, V., Celeste, A.J., Mitsock, L.M.,
Whitters, M.J., Driz, R.W., Hewick, R.M. and Wang, E.A.
(1988) Novel regulators of bone formation: molecular
clones and activities. Science 242:1528-1534.

10 Yamada, T., Placzek, M., Tanaka, H., Dodd, J. and
Jessell, T.M. (1991) Control of cell pattern in the
developing nervous system: polarizing activity of the
floor plate and notochord. Cell 64:635-647.

15 Yamada, T., Pfaff, S., Edlund, T. and Jessell, T.M.
(1993) Control of cell pattern in the neural tube: Motor
neuron induction by diffusible factors from notochord and
floor plate. Cell Submitted

-63-

SEQUENCE LISTING

(1) GENERAL INFORMATION:

5

(i) APPLICANT: Jessell, Thomas M.
Basler, Konrad
Yomada, Toshiya

10

(ii) TITLE OF INVENTION: CLONING, EXPRESSION AND USES OF
DORSALIN-1

15

(iii) NUMBER OF SEQUENCES: 18

15

(iv) CORRESPONDENCE ADDRESS:

20

(A) ADDRESSEE: Cooper & Dunham
(B) STREET: 30 Rockefeller Plaza
(C) CITY: New York
(D) STATE: New York
(E) COUNTRY: United States of America
(F) ZIP: 10112

(v) COMPUTER READABLE FORM:

25

(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

30

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:

35

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: White, John P.
(B) REGISTRATION NUMBER: 28,678
(C) REFERENCE/DOCKET NUMBER: 0576/40314

40

(ix) TELECOMMUNICATION INFORMATION:

(A) TELEPHONE: (212) 977-9550
(B) TELEFAX: (212) 664-0525
(C) TELEX: 422523 COOP UI

45

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

50

(A) LENGTH: 1603 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

55

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

60

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 91..1371

-64-

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

| | | |
|----|--|-----|
| 5 | CCTTTCTCT GTCTGTAAAG ATTCAACATT TTTAACAGT TAAAATACCTT TGTCCCTTG | 60 |
| | TCTCTCCATC AGAAAGTAAA TACATAAGAA ATG CAT TAT TTT GGA GTA TTA GCT | 114 |
| | Met His Tyr Phe Gly Val Leu Ala | |
| | 1 5 | |
| 10 | GCA CTG TCT GTT TTC AAT ATC ATT GCC TGC CTG ACA AGA GGC AAG CCT | 162 |
| | Ala Leu Ser Val Phe Asn Ile Ile Ala Cys Leu Thr Arg Gly Lys Pro | |
| | 10 15 20 | |
| 15 | TTG GAA AAC TGG AAA AAG CTA CCA GTT ATG GAA GAG TCT GAT GCA TTC | 210 |
| | Leu Glu Asn Trp Lys Lys Leu Pro Val Met Glu Glu Ser Asp Ala Phe | |
| | 25 30 35 40 | |
| 20 | TTT CAT GAT CCT GGG GAA GTG GAA CAT GAC ACC CAC TTT GAC TTT AAA | 258 |
| | Phe His Asp Pro Gly Glu Val Glu His Asp Thr His Phe Asp Phe Lys | |
| | 45 50 55 | |
| | TCT TTC TTG GAG AAT ATG AAG ACA GAT TTA CTA AGA AGT CTG AAT TTA | 306 |
| | Ser Phe Leu Glu Asn Met Lys Thr Asp Leu Leu Arg Ser Leu Asn Leu | |
| | 60 65 70 | |
| 25 | TCA AGG GTC CCC TCA CAA GTG AAG ACC AAA GAA GAG CCA CCA CAG TTC | 354 |
| | Ser Arg Val Pro Ser Gln Val Lys Thr Lys Glu Glu Pro Pro Gln Phe | |
| | 75 80 85 | |
| 30 | ATG ATT GAT TTA TAC AAC AGA TAT ACA GCG GAC AAG TCC TCC ATC CCT | 402 |
| | Met Ile Asp Leu Tyr Asn Arg Tyr Thr Ala Asp Lys Ser Ser Ile Pro | |
| | 90 95 100 | |
| 35 | GCA TCC AAC ATC GTG AGG ACC TTC AGC ACT GAA GAT GTT GTT TCT TTA | 450 |
| | Ala Ser Asn Ile Val Arg Ser Phe Ser Thr Glu Asp Val Val Ser Leu | |
| | 105 110 115 120 | |
| 40 | ATT TCA CCA GAA CAA CAC TCA TTT CAG AAA CAC ATC TTG CTC TTC AAC | 498 |
| | Ile Ser Pro Glu Glu His Ser Phe Gln Lys His Ile Leu Leu Phe Asn | |
| | 125 130 135 | |
| | ATC TCT ATT CCA CGA TAT GAG GAA GTC ACC AGA CCT GAA CTC AGA ATC | 546 |
| | Ile Ser Ile Pro Arg Tyr Glu Glu Val Thr Arg Ala Glu Leu Arg Ile | |
| | 140 145 150 | |
| 45 | TTT ATC TCC TGT CAC AAG GAA GTT GGG TCT CCC TCC AGA CTC GAA GGC | 594 |
| | Phe Ile Ser Cys His Lys Glu Val Gly Ser Pro Ser Arg Leu Glu Gly | |
| | 155 160 165 | |
| 50 | AAC ATG GTC ATT TAT GAT GTT CTA GAT GGA GAC CAT TGG GAA AAC AAA | 642 |
| | Asn Met Val Ile Tyr Asp Val Leu Asp Gly Asp His Trp Glu Asn Lys | |
| | 170 175 180 | |
| 55 | GAA AGT ACC AAA TCT TTA CTT GTC TCT CAC AGT ATT CAG GAC TGT GGC | 690 |
| | Glu Ser Thr Lys Ser Leu Leu Val Ser His Ser Ile Gln Asp Cys Gly | |
| | 185 190 195 200 | |
| 60 | TGG GAG ATG TTT GAG GTG TCC ACC GCT GTG AAA AGA TGG GTC AAG GCA | 738 |
| | Trp Glu Met Phe Glu Val Ser Ser Ala Val Lys Arg Trp Val Lys Ala | |
| | 205 210 215 | |
| | GAC AAG ATG AAG ACT AAA AAC AAG CTA GAG GTT GTT ATA GAG AGT AAG | 786 |

-65-

| | | |
|----|--|------|
| | Asp Lys Met Lys Thr Lys Asn Lys Leu Glu Val Val Ile Glu Ser Lys | |
| | 220 225 230 | |
| 5 | GAT CTG AGT GGT TTT CCT TGT GGG AAG CTG GAT ATT ACT GTT ACT CAT Asp Leu Ser Gly Phe Pro Cys Gly Lys Leu Asp Ile Thr Val Thr His | 834 |
| | 235 240 245 | |
| 10 | GAC ACT AAA AAT CTG CCC CTA TTA ATA GTG TTC TCC AAT GAT CGC AGC Asp Thr Lys Asn Leu Pro Leu Leu Ile Val Phe Ser Asn Asp Arg Ser | 882 |
| | 250 255 260 | |
| | AAT GGG ACA AAA GAG ACC AAA GTG GAG CTC CGG GAG ATG ATT GTT CAT Asn Gly Thr Lys Glu Thr Lys Val Glu Leu Arg Glu Met Ile Val His | 930 |
| | 265 270 275 280 | |
| 15 | GAA CAA GAA AGT GTG CTA AAC AAA TTA GGA AAG AAC GAC TCT TCA TCT Glu Gln Glu Ser Val Leu Asn Lys Leu Gly Lys Asn Asp Ser Ser Ser | 978 |
| | 285 290 295 | |
| 20 | GAA GAA GAA CAG AGA GAA AAA GCC ATT GCT AGG CCC CGT CAG CAT Glu Glu Glu Gln Arg Glu Glu Lys Ala Ile Ala Arg Pro Arg Gln His | 1026 |
| | 300 305 310 | |
| 25 | TCC TCC AGA AGC AAG AGA AGC ATA GGA GCA AAC CAC TGT CGG AGA ACG Ser Ser Arg Ser Lys Arg Ser Ile Gly Ala Asn His Cys Arg Arg Thr | 1074 |
| | 315 320 325 | |
| 30 | TCA CTC CAT GTG AAC TTT AAA GAA ATA GGT TGG GAT TCT TGG ATC ATT Ser Leu His Val Asn Phe Lys Glu Ile Gly Trp Asp Ser Trp Ile Ile | 1122 |
| | 330 335 340 | |
| | GCA CCC AAA GAT TAT GAG GCT TTT GAG TGT AAA GGA GGT TGC TTC TTC Ala Pro Lys Asp Tyr Glu Ala Phe Glu Cys Lys Gly Gly Cys Phe Phe | 1170 |
| | 345 350 355 360 | |
| 35 | CCC CTC ACA GAT AAT GTT ACG CCA ACC AAA CAT GCT ATT GTC CAG ACT Pro Leu Thr Asp Asn Val Thr Pro Thr Lys His Ala Ile Val Gln Thr | 1218 |
| | 365 370 375 | |
| 40 | CTG GTG CAT CTC CAA AAC CCA AAG AAA GCT TCC AAG GCC TGT TGT GTT Leu Val His Leu Gln Asn Pro Lys Lys Ala Ser Lys Ala Cys Cys Val | 1266 |
| | 380 385 390 | |
| 45 | CCA ACT AAA TTG GAT GCA ATC TCT ATT CTT TAT AAG GAT GAT GCT GGT Pro Thr Lys Leu Asp Ala Ile Ser Ile Leu Tyr Lys Asp Asp Ala Gly | 1314 |
| | 395 400 405 | |
| 50 | GTG CCC ACT TTG ATA TAT AAC TAT GAA GGG ATG AAA GTG GCA GAA TGT Val Pro Thr Leu Ile Tyr Asn Tyr Glu Gly Met Lys Val Ala Glu Cys | 1362 |
| | 410 415 420 | |
| | GGC TGC AGG TAGTATATGC TGAATATCTA AGAATATACT CTTTCTGCT | 1411 |
| | Gly Cys Arg | |
| | 425 | |
| 55 | GTCTGTGAAA CTGTACATTA GTGATGCAAA TGAAATCCT TGCAAAACAAG GTTTGGAGCA | 1471 |
| | CGGCATGGGG CTGGTTGTTG TTGCTGCTTT TAAAGGAAAG ATGCCATTAA AAGAATGGCA | 1531 |
| 60 | ATCACTGTAA ATACCCCTGCA TTATATACCA TTAATTAAGA CTTTGTGAGA TTGAAAAAAA | 1591 |
| | AAAAAAAAAA AA | 1603 |

-66-

(2) INFORMATION FOR SEQ ID NO:2:

5 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 427 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: protein

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met His Tyr Phe Gly Val Leu Ala Ala Leu Ser Val Phe Asn Ile Ile
1 5 10 15

15 Ala Cys Leu Thr Arg Gly Lys Pro Leu Glu Asn Trp Lys Lys Leu Pro
20 25 30

20 Val Met Glu Glu Ser Asp Ala Phe Phe His Asp Pro Gly Glu Val Glu
35 40 45

His Asp Thr His Phe Asp Phe Lys Ser Phe Leu Glu Asn Met Lys Thr
50 55 60

25 Asp Leu Leu Arg Ser Leu Asn Leu Ser Arg Val Pro Ser Gln Val Lys
65 70 75 80

30 Thr Lys Glu Glu Pro Pro Gln Phe Met Ile Asp Leu Tyr Asn Arg Tyr
85 90 95

35 Thr Ala Asp Lys Ser Ser Ile Pro Ala Ser Asn Ile Val Arg Ser Phe
100 105 110

40 Ser Thr Glu Asp Val Val Ser Leu Ile Ser Pro Glu Glu His Ser Phe
115 120 125

45 Gln Lys His Ile Leu Leu Phe Asn Ile Ser Ile Pro Arg Tyr Glu Glu
130 135 140

50 Val Thr Arg Ala Glu Leu Arg Ile Phe Ile Ser Cys His Lys Glu Val
145 150 155 160

55 Gly Ser Pro Ser Arg Leu Glu Gly Asn Met Val Ile Tyr Asp Val Leu
165 170 175

60 Asp Gly Asp His Trp Glu Asn Lys Glu Ser Thr Lys Ser Leu Leu Val
180 185 190

65 Ser His Ser Ile Gln Asp Cys Gly Trp Glu Met Phe Glu Val Ser Ser
195 200 205

70 Ala Val Lys Arg Trp Val Lys Ala Asp Lys Met Lys Thr Lys Asn Lys
210 215 220

75 Leu Glu Val Val Ile Glu Ser Lys Asp Leu Ser Gly Phe Pro Cys Gly
225 230 235 240

80 Lys Leu Asp Ile Thr Val Thr His Asp Thr Lys Asn Leu Pro Leu Leu
245 250 255

85 Ile Val Phe Ser Asn Asp Arg Ser Asn Gly Thr Lys Glu Thr Lys Val
260 265 270

-67-

Glu Leu Arg Glu Met Ile Val His Glu Gln Glu Ser Val Leu Asn Lys
 275 280 285
 5 Leu Gly Lys Asn Asp Ser Ser Ser Glu Glu Glu Gln Arg Glu Glu Lys
 290 295 300
 Ala Ile Ala Arg Pro Arg Gln His Ser Ser Arg Ser Lys Arg Ser Ile
 305 310 315 320
 10 Gly Ala Asn His Cys Arg Arg Thr Ser Leu His Val Asn Phe Lys Glu
 325 330 335
 Ile Gly Trp Asp Ser Trp Ile Ile Ala Pro Lys Asp Tyr Glu Ala Phe
 340 345 350
 15 Glu Cys Lys Gly Gly Cys Phe Phe Pro Leu Thr Asp Asn Val Thr Pro
 355 360 365
 20 Thr Lys His Ala Ile Val Gln Thr Leu Val His Leu Gln Asn Pro Lys
 370 375 380
 Lys Ala Ser Lys Ala Cys Cys Val Pro Thr Lys Leu Asp Ala Ile Ser
 385 390 395 400
 25 Ile Leu Tyr Lys Asp Asp Ala Gly Val Pro Thr Leu Ile Tyr Asn Tyr
 405 410 415
 Glu Gly Met Lys Val Ala Glu Cys Gly Cys Arg
 420 425
 30 (2) INFORMATION FOR SEQ ID NO:3:
 (i) SEQUENCE CHARACTERISTICS:
 35 (A) LENGTH: 143 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: protein
 40 (iii) HYPOTHETICAL: NO
 (iv) ANTI-SENSE: NO
 45 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:
 Glu His Ser Trp Ser Gln Ile Arg Pro Leu Leu Val Thr Phe Gly His
 50 1 5 10 15
 Asp Gly Lys Gly His Pro Leu His Lys Arg Glu Lys Arg Gln Ala Lys
 55 20 25 30
 His Lys Gln Arg Lys Arg Leu Lys Ser Ser Cys Lys Arg His Pro Leu
 35 40 45
 Tyr Val Asp Phe Ser Asp Val Gly Trp Asn Asp Trp Ile Val Ala Pro
 60 50 55 60
 65 Pro Gly Tyr His Ala Phe Tyr Cys His Gly Glu Cys Pro Phe Pro Leu
 70 75 80

-68-

Ala Asp His Leu Asn Ser Thr Asn His Ala Ile Val Gln Thr Leu Val
85 90 95
Asn Ser Val Asn Ser Lys Ile Pro Lys Ala Cys Cys Val Pro Thr Glu
5 100 105 110
Leu Ser Ala Ile Ser Met Leu Tyr Leu Asp Glu Asn Glu Lys Val Val
115 120 125
10 Leu Lys Asn Tyr Gln Asp Met Val Val Glu Gly Cys Gly Cys Arg
130 135 140

(2) INFORMATION FOR SEQ ID NO:4:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 144 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
20 (ii) MOLECULE TYPE: protein
(iii) HYPOTHETICAL: NO
25 (iv) ANTI-SENSE: NO

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:
Asp Asp Gly Arg His Lys Ala Arg Ser Ile Arg Asp Val Ser Gly Gly
1 5 10 15
35 Glu Gly Gly Lys Gly Gly Arg Asn Lys Arg His Ala Arg Arg Pro
20 25 30
40 Thr Arg Arg Lys Asn His Asp Asp Thr Cys Arg Arg His Ser Leu Tyr
35 40 45
45 Val Asp Phe Ser Asp Val Gly Trp Asp Asp Trp Ile Val Ala Pro Leu
50 55 60
55 Gly Tyr Asp Ala Tyr Tyr Cys His Gly Lys Cys Pro Phe Pro Leu Ala
65 70 75 80
45 Asp His Phe Asn Ser Thr Asn His Ala Val Val Gln Thr Leu Val Ala
85 90 95
50 Asn Asn Met Asn Pro Gly Lys Val Pro Lys Ala Cys Cys Val Pro Thr
100 105 110
55 Gln Leu Asp Ser Val Ala Met Leu Tyr Leu Asn Asp Gln Ser Thr Val
115 120 125
55 Val Leu Lys Asn Tyr Gln Glu Met Thr Val Val Gly Cys Gly Cys Arg
130 135 140

(2) INFORMATION FOR SEQ ID NO:5:

60 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 143 amino acids

-69-

- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

10 (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

15 Arg Thr Thr Arg Ser Ala Ser Ser Arg Arg Arg Gln Gln Ser Arg Asn
1 5 10 15

20 Arg Ser Thr Gln Ser Gln Asp Val Ala Arg Val Ser Ser Ala Ser Asp
20 25 30

25 Tyr Asn Ser Ser Glu Leu Lys Thr Ala Cys Arg Lys His Glu Leu Tyr
35 40 45

30 Val Ser Phe Gln Asp Leu Gly Trp Gln Asp Trp Ile Ile Ala Pro Lys
50 55 60

35 Gly Tyr Ala Ala Asn Tyr Cys Asp Gly Glu Cys Ser Phe Pro Leu Asn
65 70 75 80

40 Ala His Met Asn Ala Thr Asn His Ala Ile Val Gln Thr Leu Val His
85 90 95

45 Leu Met Asn Pro Glu Tyr Val Pro Lys Pro Cys Cys Ala Pro Thr Lys
100 105 110

50 Leu Asn Ala Ile Ser Val Leu Tyr Phe Asp Asp Asn Ser Asn Val Ile
115 120 125

55 Leu Lys Lys Tyr Arg Asn Met Val Val Arg Ala Cys Gly Cys His
130 135 140

(2) INFORMATION FOR SEQ ID NO:6:

45 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 144 amino acids
- (B) TYPE: amino acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

55 (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

60 Glu Cys Lys Asp Ile Gln Thr Phe Leu Tyr Thr Ser Leu Leu Thr Val
1 5 10 15

-70-

Thr Leu Asn Pro Leu Arg Cys Lys Arg Pro Arg Arg Lys Arg Ser Tyr
 20 25 30
 Ser Lys Leu Pro Phe Thr Ala Ser Asn Ile Cys Lys Lys Arg His Leu
 5 35 40 45
 Tyr Val Glu Phe Lys Asp Val Gly Trp Gln Asn Trp Val Ile Ala Pro
 50 55 60
 10 Gln Gly Tyr Met Ala Asn Tyr Cys Tyr Gly Glu Cys Pro Tyr Pro Leu
 65 70 75 80
 Thr Glu Ile Leu Asn Gly Ser Asn His Ala Ile Leu Gln Thr Leu Val
 15 85 90 95
 His Ser Ile Glu Pro Glu Asp Ile Pro Leu Pro Cys Cys Val Pro Thr
 100 105 110
 20 Lys Met Ser Pro Ile Ser Met Leu Phe Tyr Asp Asn Asn Asp Asn Val
 115 120 125
 Val Leu Arg His Tyr Glu Asn Met Ala Val Asp Glu Cys Gly Cys Arg
 25 130 135 140
 (2) INFORMATION FOR SEQ ID NO:7:
 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 147 amino acids
 30 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
 (ii) MOLECULE TYPE: protein
 35 (iii) HYPOTHETICAL: NO
 (iv) ANTI-SENSE: NO
 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
 Gly Ala Asp Glu Glu Lys Glu Gln Ser His Arg Pro Phe Leu Met Leu
 45 1 5 10 15
 Gln Ala Arg Gln Ser Glu Asp His Pro His Arg Arg Arg Arg Gly
 50 20 25 30
 Leu Glu Cys Asp Gly Lys Val Asn Ile Cys Cys Lys Gln Phe Phe
 55 35 40 45
 Val Ser Phe Lys Asp Ile Gly Trp Asn Asp Trp Ile Ile Ala Pro Ser
 60 50 55 60
 Gly Tyr His Ala Asn Tyr Cys Glu Gly Glu Cys Pro Ser His Ile Ala
 65 70 75 80
 Gly Thr Ser Gly Ser Ser Leu Ser Phe His Ser Thr Val Ile Asn His
 75 85 90 95
 Tyr Arg Met Arg Gly His Ser Pro Phe Ala Asn Leu Lys Ser Cys Cys

-71-

| | | | |
|----|---|-------------------------|-----|
| | 100 | 105 | 110 |
| | Val Pro Thr Lys Leu Arg Pro Met Ser Met | Leu Tyr Tyr Asp Asp Gly | |
| | 115 | 120 | 125 |
| 5 | Gln Asn Ile Ile Lys Lys Asp Ile Gln Asn Met | Ile Val Glu Glu Cys | |
| | 130 | 135 | 140 |
| 10 | Gly Cys Ser | | |
| | 145 | | |

(2) INFORMATION FOR SEQ ID NO:8:

15 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 139 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

25 (iv) ANTI-SENSE: NO

30 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

| | | | |
|---|-----|-----|----|
| Gly Met Asn Arg Pro Phe Leu Leu Leu Met Ala Thr Pro Leu Glu Arg | | | |
| 1 | 5 | 10 | 15 |
| Ala Gln His Leu Gln Ser Ser Arg His Arg Arg Ala Leu Asp Thr Asn | | | |
| 20 | 25 | 30 | |
| 35 Tyr Cys Phe Ser Ser Thr Glu Lys Asn Cys Cys Val Arg Gln Leu Tyr | | | |
| 35 | 40 | 45 | |
| 40 Ile Asp Phe Arg Lys Asp Leu Gly Trp Lys Trp Ile His Glu Pro Lys | | | |
| 50 | 55 | 60 | |
| 45 Gly Tyr His Ala Asn Phe Cys Leu Gly Pro Cys Pro Tyr Ile Trp Ser | | | |
| 65 | 70 | 75 | 80 |
| 55 Leu Asp Thr Gln Tyr Ser Lys Val Leu Ala Leu Tyr Asn Gln His Asn | | | |
| 85 | 90 | 95 | |
| 50 Pro Gly Ala Ser Ala Ala Pro Cys Cys Val Pro Gln Ala Leu Glu Pro | | | |
| 100 | 105 | 110 | |
| 60 55 Leu Pro Ile Val Tyr Tyr Val Gly Arg Lys Pro Lys Val Glu Gln Leu | | | |
| 115 | 120 | 125 | |
| 55 Ser Asn Met Ile Val Arg Ser Cys Lys Cys Ser | | | |
| 130 | 135 | | |

(2) INFORMATION FOR SEQ ID NO:9:

60 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 257 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single

-72-

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

5 (iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Asp Val Leu Glu Asp Ser Glu Thr Trp Asp Gln Ala Thr Gly Thr Lys
1 5 10 1515 Thr Phe Leu Val Ser Gln Asp Ile Arg Asp Glu Gly Trp Glu Thr Leu
20 25 3020 Glu Val Ser Ser Ala Val Lys Arg Trp Val Arg Ala Asp Ser Thr Thr
35 40 45Asn Lys Asn Lys Leu Glu Val Thr Val Gln Ser His Arg Glu Ser Cys
50 55 6025 Asp Thr Leu Asp Ile Ser Val Pro Pro Gly Ser Lys Asn Leu Pro Phe
65 70 75 80Phe Val Val Phe Ser Asn Asp Arg Ser Asn Gly Thr Lys Glu Thr Arg
85 90 9530 Leu Asp Leu Leu Lys Glu Met Ile Gly His Glu Gln Glu Thr Met Leu
100 105 11035 Val Lys Thr Ala Lys Asn Ala Tyr Gln Gly Ala Gly Glu Ser Gln Glu
115 120 125Glu Glu Gly Leu Asp Gly Tyr Thr Ala Val Gly Pro Leu Leu Ala Arg
130 135 14040 Arg Lys Arg Ser Thr Gly Ala Ser Ser His Cys Gln Lys Thr Ser Leu
145 150 155 160Arg Val Asn Phe Glu Asp Ile Gly Trp Asp Ser Trp Ile Ile Ala Pro
165 170 17545 Lys Glu Tyr Asp Ala Tyr Glu Cys Lys Gly Gly Cys Phe Phe Pro Leu
180 185 19050 Ala Asp Asp Val Thr Pro Thr Lys His Ala Ile Val Gln Thr Leu Val
195 200 205His Leu Lys Phe Pro Thr Lys Val Gly Lys Ala Cys Cys Val Pro Thr
210 215 22055 Lys Leu Ser Pro Ile Ser Ile Leu Tyr Lys Asp Asp Met Gly Val Pro
225 230 235 240Thr Leu Lys Tyr His Tyr Glu Gly Met Ser Val Ala Glu Cys Gly Cys
245 250 255

60 Arg

-73-

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

5

(ii) MOLECULE TYPE: cDNA

10

(iii) HYPOTHETICAL: YES

15

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TGGAATTCTG GVANGAYTGG ATHRTNGC

28

20

(2) INFORMATION FOR SEQ ID NO:11:

25

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 28 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

30

(iii) HYPOTHETICAL: YES

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

35

GAGGATCCAR NGTYTGNACD ATNGCRTG

28

(2) INFORMATION FOR SEQ ID NO:12:

40

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 29 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

45

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: YES

50

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

TGGAATTCAT CGATAACCGGA AGCTGAAGC

29

55

(2) INFORMATION FOR SEQ ID NO:13:

60

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 32 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

-74-

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: YES

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

AGCGTCCGACA TCGATATTCA GCATATACTA CC

10

(2) INFORMATION FOR SEQ ID NO:14:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 45 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: YES

25

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

CGGAATTCTGA TATCAGCTTC TGCTCTGCTC CTATGCTTCT CTTGC

45

30 (2) INFORMATION FOR SEQ ID NO:15:

35 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 47 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

40 (iii) HYPOTHETICAL: YES

45

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

CGGAATTCTGA TATCCGAGGA GGACCTGAAC CACTGTCGGA GAACGTC

47

(2) INFORMATION FOR SEQ ID NO:16:

50 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

55 (ii) MOLECULE TYPE: protein

60

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Ser Ile Gly Ala Glu Gln Lys Leu Ile Ser

-75-

1 5 10

(2) INFORMATION FOR SEQ ID NO:17:

5 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 4 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: protein

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Arg Ser Lys Arg
1

20 (2) INFORMATION FOR SEQ ID NO:18:

25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 10 amino acids
 (B) TYPE: amino acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

30 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

35 Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu
 1 5 10

-76-

What is claimed is:

1. An isolated, vertebrate nucleic acid molecule encoding dorsalin-1.
5
2. An isolated, vertebrate DNA molecule of claim 1.
10
3. An isolated, vertebrate cDNA molecule of claim 2.
15
4. An isolated, vertebrate genomic DNA molecule of claim 2.
20
5. An isolated, vertebrate RNA molecule of claim 1.
25
6. An isolated, human nucleic acid molecule of claim 1.
30
7. An isolated, mouse nucleic acid molecule of claim 1.
35
8. An isolated, chick nucleic acid molecule of claim 1.
40
9. A nucleic acid molecule comprising a nucleic acid molecule of at least 15 nucleotides capable of specifically hybridizing with a nucleic acid molecule of claim 1.
45
10. An isolated nucleic acid molecule of claim 2 operatively linked to a promoter of RNA transcription.
50

-77-

11. A vector which comprises the isolated nucleic acid molecule of claim 10.
12. A vector of claim 10, wherein the isolated nucleic acid molecule is linked to a plasmid.
- 5 13. The plasmid of claim 12 designated pKB502 (ATCC Accession No. 75321).
- 10 14. A host vector system for the production of a polypeptide having the biological activity of dorsalin-1 which comprises the vector of claim 11 in a suitable host.
- 15 15. A host vector system of claim 14, wherein the suitable host is a bacterial cell, insect cell, or animal cell.
- 20 16. A method of producing a polypeptide having the biological activity of dorsalin-1 which comprises growing the host vector system of claim 14 under suitable conditions permitting production of the polypeptide and recovering the polypeptide so produced.
- 25 17. A purified vertebrate dorsalin-1.
18. A purified human dorsalin-1 of claim 17.
- 30 19. A polypeptide encoded by the isolated vertebrate nucleic acid molecule of claim 1.
20. A method for stimulating neural crest cell differentiation in a culture comprising administering an amount of the purified
- 35

-78-

dorsalin-1 of claim 17 effective to stimulate neural crest cell differentiation to the culture.

- 5 21. A method for stimulating neural crest cell differentiation in a subject comprising administering to the subject an amount of the purified dorsalin-1 of claim 17 effective to stimulate neural crest cell differentiation.
- 10 22. A method for regenerating nerve cells in a subject comprising administering to the subject an amount of the purified dorsalin-1 of claim 17 effective to regenerate nerve cells.
- 15 23. A method for promoting bone growth in a subject comprising administering to the subject an amount of the purified dorsalin-1 of claim 17 effective to promote bone growth.
- 20 24. A method for promoting wound healing in a subject comprising administering to the subject an amount of the purified dorsalin-1 of claim 17 effective to promote wound healing.
- 25 25. A method for treating neural tumor in a subject comprising administering to the subject an amount of the purified dorsalin-1 of claim 17 effective to inhibit the tumor cell growth.
- 30 26. A method of claim 25, wherein the neural tumor is neurofibroma.
- 35 27. A method of claim 25, wherein the neural tumor is Schwann cell tumor.

-79-

28. A pharmaceutical composition for stimulating neural crest cell differentiation comprising an amount of the purified dorsalin-1 of claim 17 effective to stimulate neural crest cell differentiation and a pharmaceutically acceptable carrier.
29. A pharmaceutical composition for regenerating nerve cells in a subject comprising an amount of the purified dorsalin-1 of claim 17 effective to regenerate nerve cells and a pharmaceutically acceptable carrier.
30. A pharmaceutical composition for promoting bone growth in a subject comprising an amount of the purified dorsalin-1 of claim 17 effective to promote bone growth and a pharmaceutically acceptable carrier.
31. A pharmaceutical composition for promoting wound healing in a subject comprising an amount of the purified dorsalin-1 of claim 17 effective to promote wound healing and a pharmaceutically acceptable carrier.
32. A pharmaceutical composition for treating neural tumor in a subject comprising an amount of the purified dorsalin-1 of claim 17 effective to inhibit neural tumor cell growth and a pharmaceutically acceptable carrier.
33. A pharmaceutical composition of claim 32, wherein the neural tumor is neurofibroma.
34. A pharmaceutical composition of claim 33,

-80-

wherein the neural tumor is Schwann cell tumor.

35. A method to produce antibody using the purified dorsalin-1 of claim 18.

5

36. Antibody capable of binding to dorsalin-1.

37. A monoclonal antibody of claim 36.

10 38. An antibody of claim 36 capable of inhibiting the biological activity of dorsalin-1.

15 39. A method for inhibiting dorsalin-1 activity in a subject comprising administering to the subject an amount of the antibody of claim 38 effective to inhibit the dorsalin-1 activity.

20 40. A pharmaceutical composition for inhibiting dorsalin-1 activity comprising an amount of antibody of claim 38 effective to inhibit dorsalin-1 activity and a pharmaceutically acceptable carrier.

FIGURE 1A

CCTTCCCTGCTGTAAGATTCAACATTAAATCAGTTAAATCAGTTAAACATTCAGTTAAATCAGTAATCAGAAAGTAATACATAAGAA
 M H Y F G V L A A L S V F N I I A C L T R G K P L E N W K K 30
 ATGCATTATTGGAGTATTAGCTGCACTGCTGTTCAATTATCATTGCCTGACAAGAGGCAAGGCCCTTGGAAAACCTGGAAAAAG
 L P V M E E S D A F F H D P G E V E H D T H F D F K S F L E 60
 CTACCACTTATGGAAGAGTCTGATGGCATTCTTCAATGATCCTGGGGAAAGTGGAAACATGACACCCACTTGAACTTAAATCTTCTTGAG
 N M K T D L L R S L N L S R V P S Q V K T K E E P P Q F M I 90
 ATATGAAGACAGATTACTAAGAAGTCTGAATTATCAAGGGTCCCTCACAAAGTGAAGACCAAAAGAGCCACACAGTTCATGATT
 D L Y N R Y T A D K S S I P A S N I V R S F S T E D V V S L 120
 GATTATACACAGATATAACAGGGACAAAGTCCCTCAATCCMACATCGTGAAGGAGCTTCAGCACTGAAGATGTTGTTCTTTA
 I S P E E H S F Q K H I L L F N I S I P R Y E E V T R A E L 150
 ATTCACCAAGAAACACTCATTCAGAAACATTCAGAAACATCTTGCTCTTCAACATCTCTATTCCACCGATATGAGGAAGTCAACAGGCTGAACCTG
 R I F I S C H K E V G S P S R L E G N M V I Y D V L D G D H 180
 AGAATCTTATCTCCTGTCACAAGGAAGTGGCTCCAGACTGGAAAGCCACATGGTCATTATGATGTTCTAGATGGAGACCAT
 W E N K E S T K S L L V S H S I Q D C G W E M F E V S S A V 210
 TGGGAAACAAAGAACATCCAAATCTTACTTGTCTCACAGTATTCAAGGACTGTTGAGGAGATGTTGAGGTGTCAGGGCTGCTGTG
 K R W V K A D K M K T K N K L E V I E S K D L S G F P C G 240
 AAAAGATGGGTCAAGGGAGACAAGAGTGAAGACTAAACAAAGCTAGAGGTGTTATAGAGAGTAAGGATCTGAGTGGTTTCCTGTTGGG

SUBSTITUTE SHEET (RULE 26)

1/16

FIGURE 1B

3/16

FIGURE 2A

| | | | | | | |
|------------|--|-----|-----|-----------|---------|-------|
| DORSALIN-1 | SVLNKLGKNDSSSEEQREEKAIAARPRQHSSRSRKR^SIGANHCRRTS^HVNFE-KEI | GWD | SWI | IAPKDYEAF | ECKX | GGCF |
| BMP-2 | EHSSWSQIRPLLVTFGHDGKGHPLHKEKRQAKHKQRKRLKSSCKRHPLYVDF | SDV | GWN | DWIVAPP | GYHAF | YCH |
| DIF | DDGRHKARSIR^DSSGEGGGGGRNIKRHARRPTTRKHNDDTCRHSLYVDF | SDV | GWD | DWIVAPL | GYDAY | YCH |
| BMP-6 | RTTRASSASSRRQQSRNRSRQQDVARVSSASDYNSSQELTA | SDV | GWD | DWIVAPK | GYAANYC | DGECS |
| VG-1 | ECKDIDQTFLYTSLLTVTLNPLRCKRPRRKRSYSKLPFTASNICKKRHLYVDF | SDV | GWD | DWIVAPQG | GYMANYC | YGECP |
| ACTIVINA | GADDEKEQSHPFLMLQARQSEDHPHRRRRR^GLECDGKVNIC | SDV | GWD | DWIVAPSG | GYHANYC | EGEC |
| TGF-BETA1 | GMRPFLLMMATPLERAQHLSRQHRR^ALDTNYCFSSTEKNQ | SDV | GWD | DWIVAPK | GYHANF | GP |

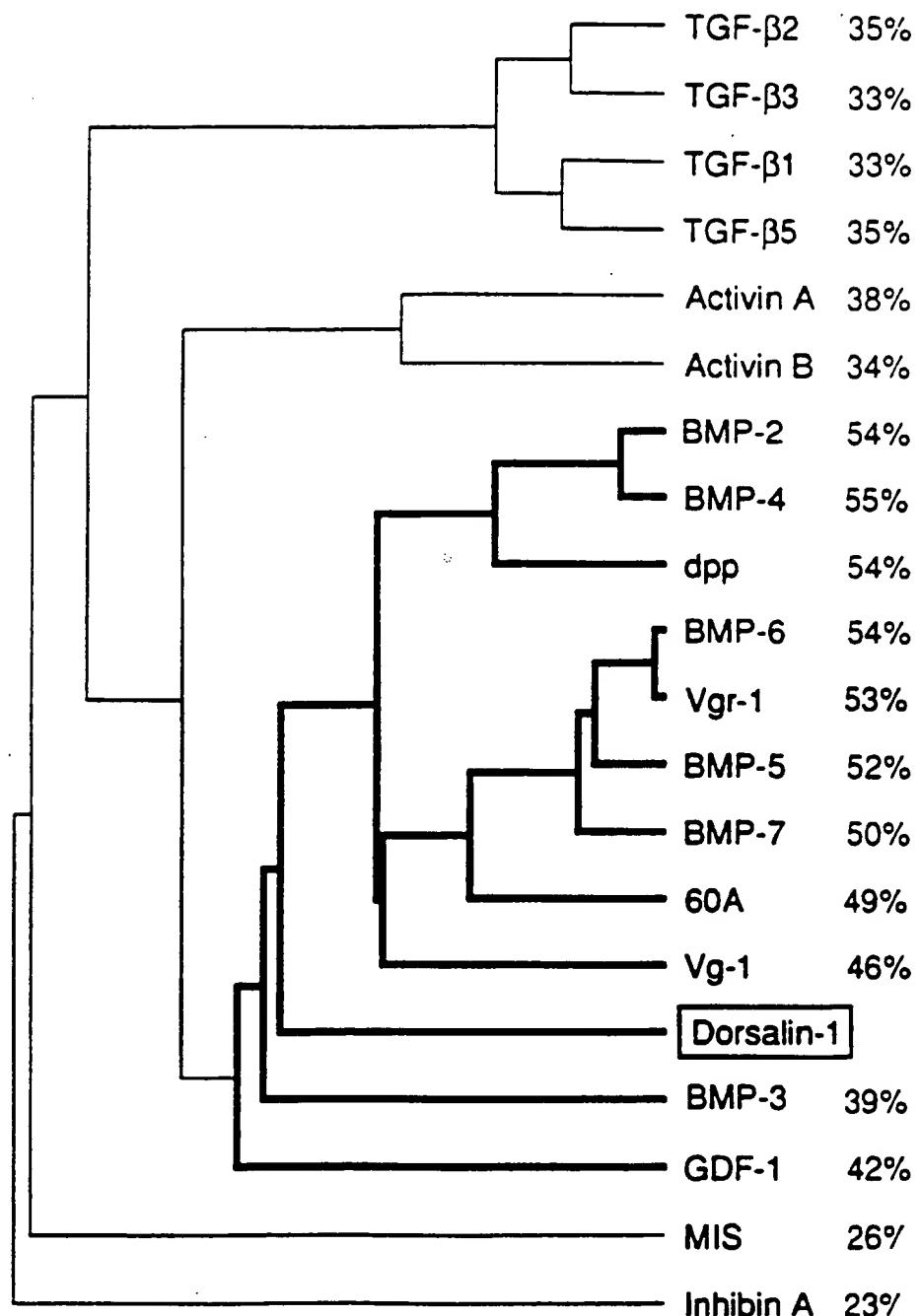
*

| | | | | | | |
|------------|-----------------------------|-------|-------------------|-------|------------|-------|
| DORSALIN-1 | FPLTDNYTPTIKHAIVQTLVHLq | ----- | NPKKASKKACCVPTKLD | ----- | DDAGVPT | ----- |
| BMP-2 | FPLADHLLNSINHAIVQTLVNSY | ----- | NPKKA | ----- | YKVAECGGCR | [427] |
| DIF | FPLADHFNSINHAIVQTLVNNH | ----- | SKIP | ----- | -NQD | [396] |
| BMP-6 | FPLNAAHMNAUNHAIVQTLVLM | ----- | ACCVPTK | ----- | YQEE | [588] |
| VG-1 | YPLTEIILGNSNHAIIQTLVHS1 | ----- | KAPTKLN | ----- | TTVGGCR | [514] |
| ACTIVINA | SHIAGTSGSSLSFHSTVINHYMRGHS | ----- | SPISM | ----- | -KRN | [360] |
| TGF-BETA1 | -----YIWSLDTQYSKVLALY-NQHNP | ----- | PTKLRPMSM | ----- | YRA | [427] |

*

4/16

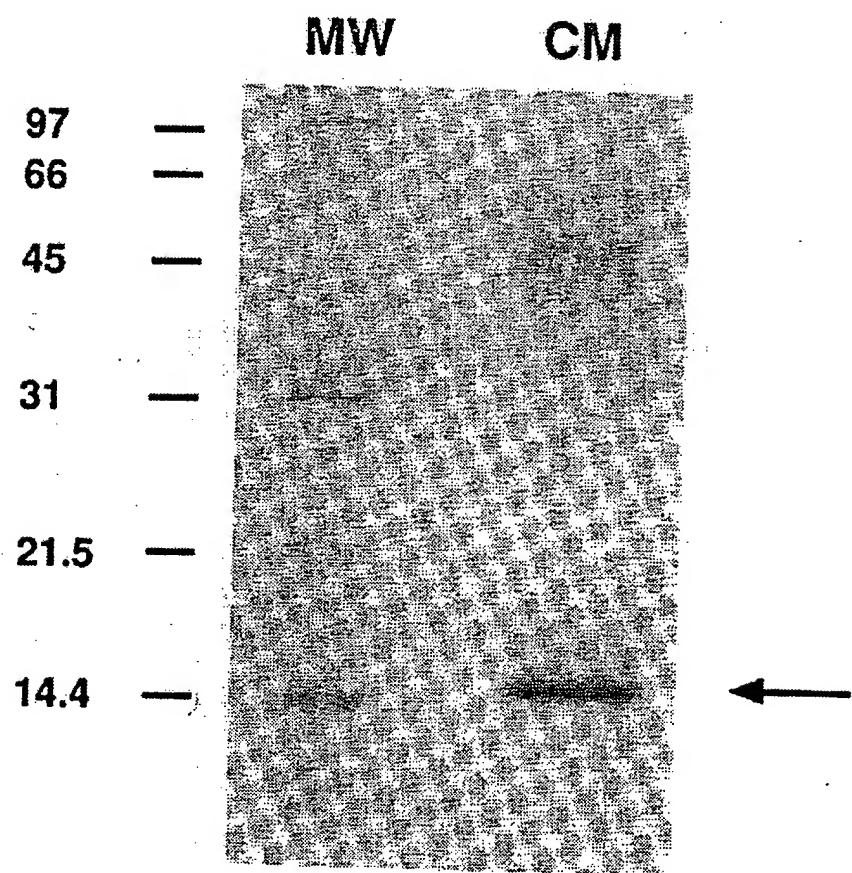
FIGURE 2B



SUBSTITUTE SHEET (RULE 26)

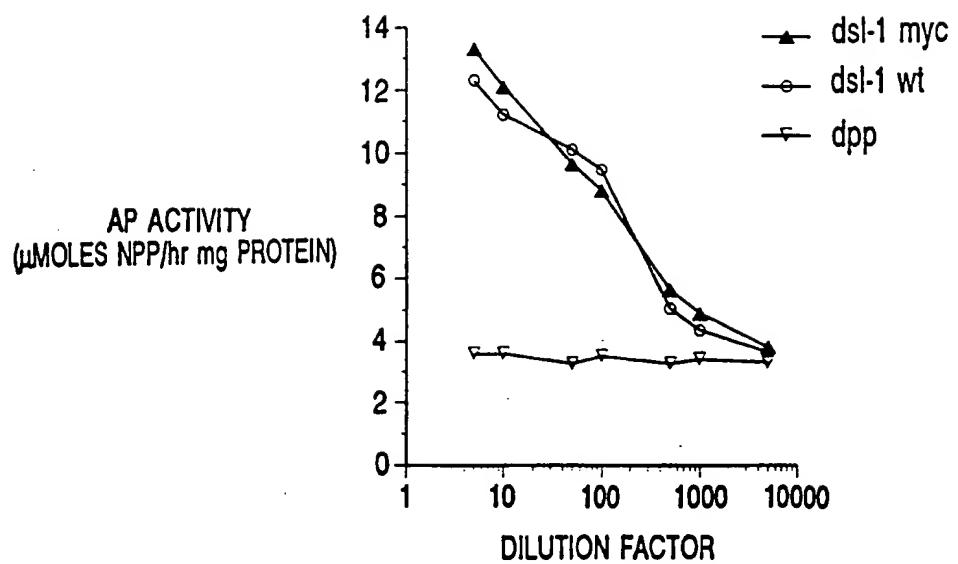
5/16

FIGURE 3A



6/16

FIGURE 3B



SUBSTITUTE SHEET (RULE 26)

7/16

FIGURE 4A FIGURE 4B FIGURE 4C FIGURE 4D

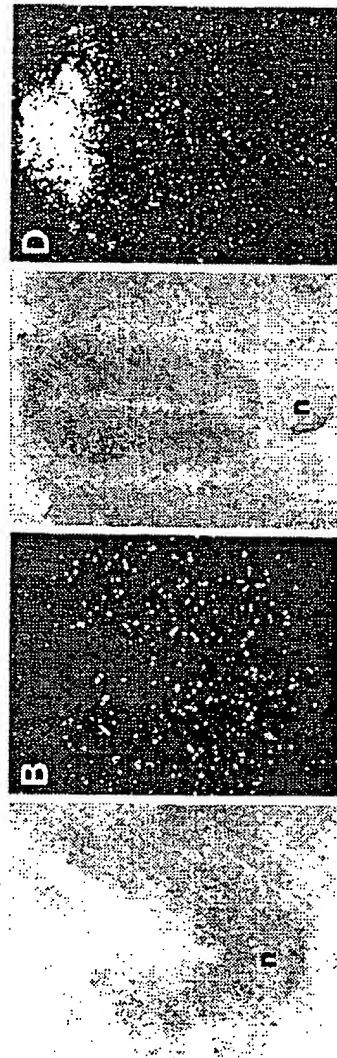
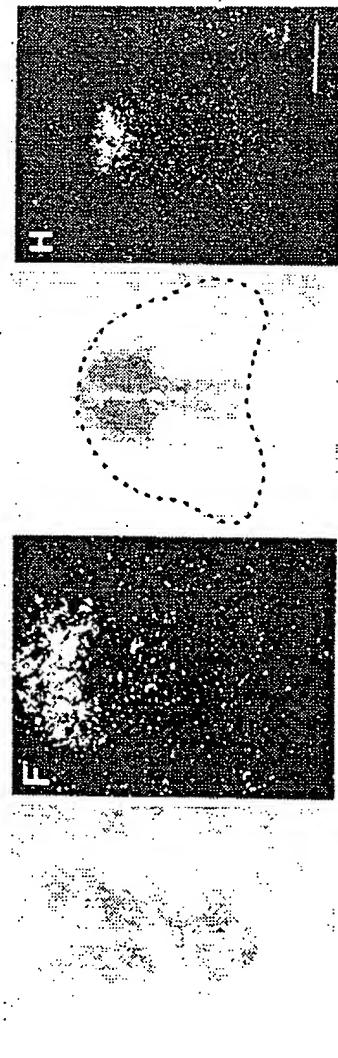
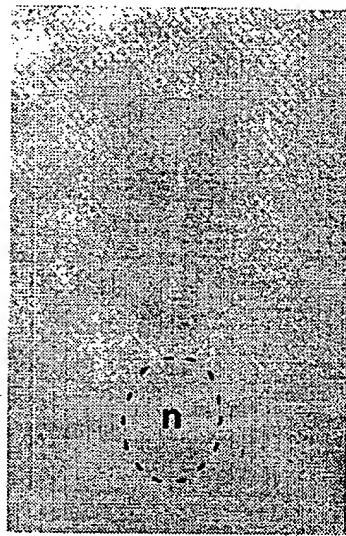
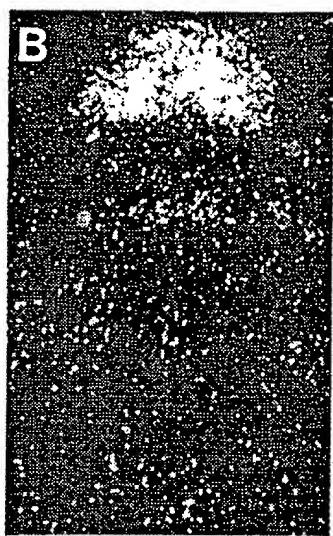
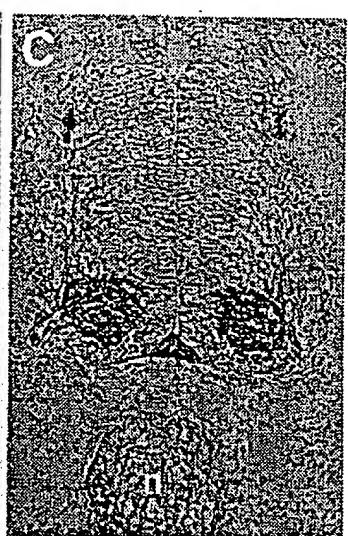
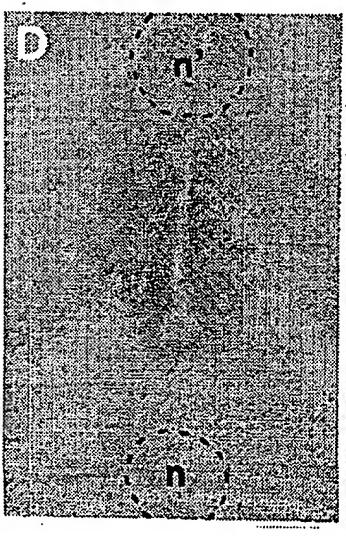
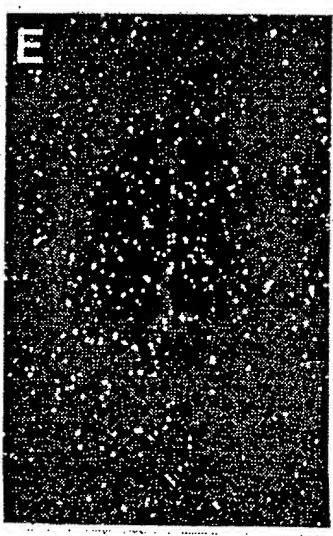
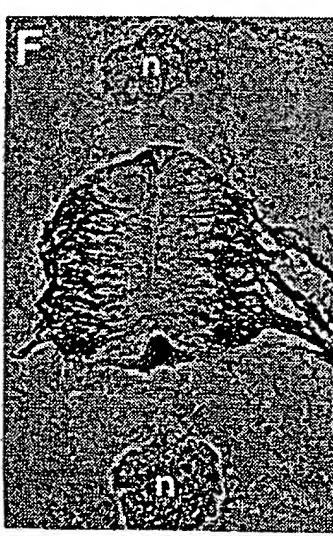


FIGURE 4E FIGURE 4F FIGURE 4G FIGURE 4H



8/16

FIGURE 5A**FIGURE 5B****FIGURE 5C****FIGURE 5D****FIGURE 5E****FIGURE 5F**

9/16

FIGURE 5G

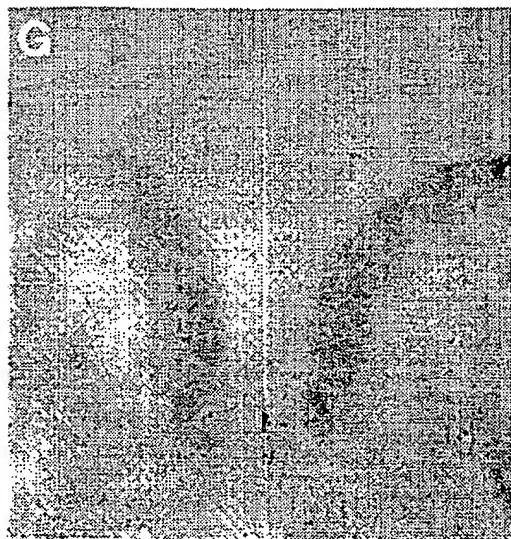
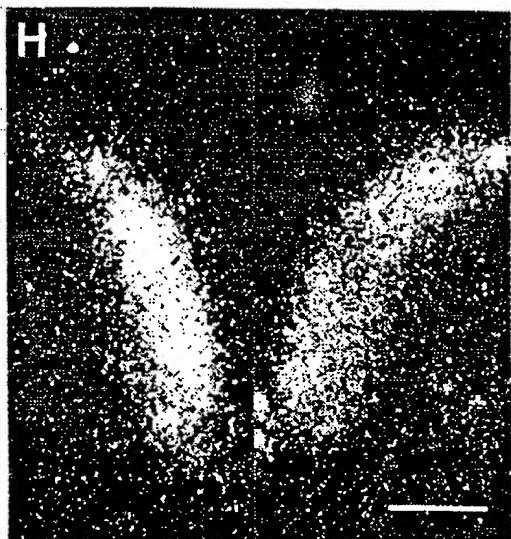


FIGURE 5H



10/16

FIGURE 6A **FIGURE 6B** **FIGURE 6C**

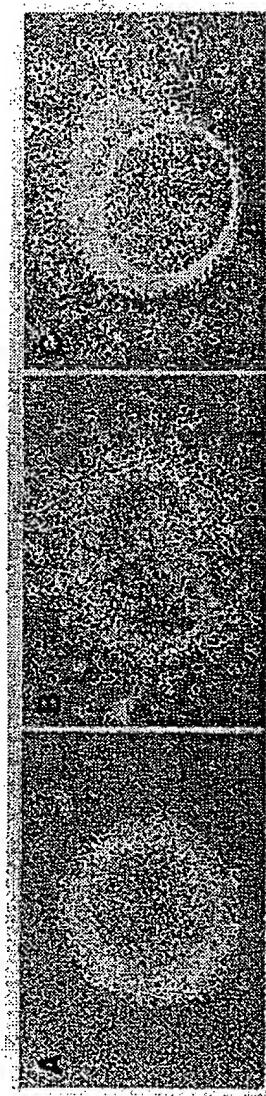
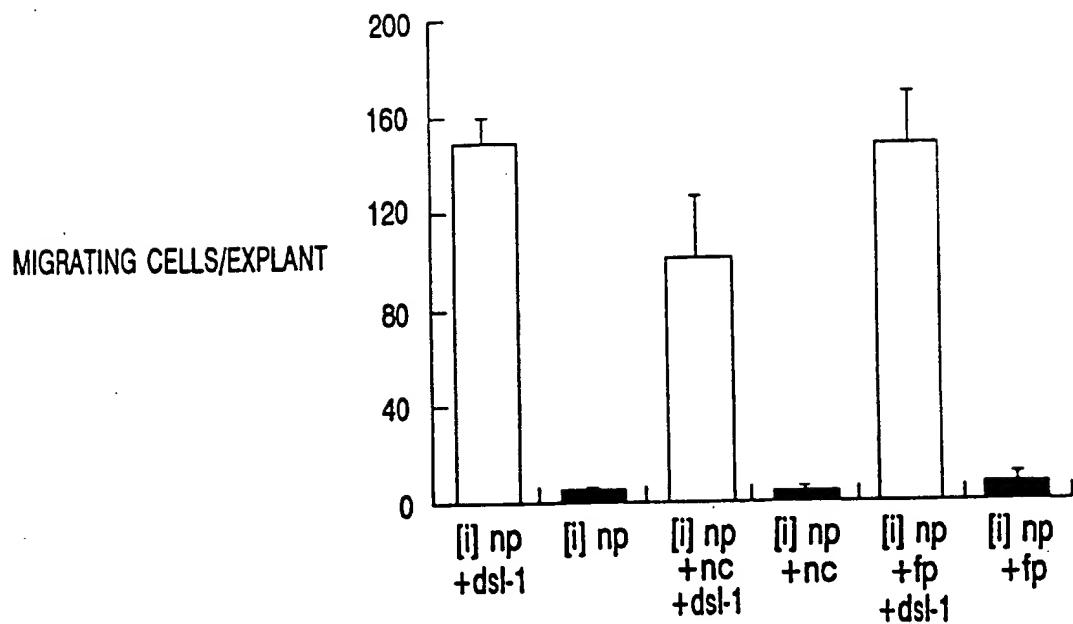


FIGURE 6D **FIGURE 6E** **FIGURE 6F**



11/16

FIGURE 6G



12/16

FIGURE 7A

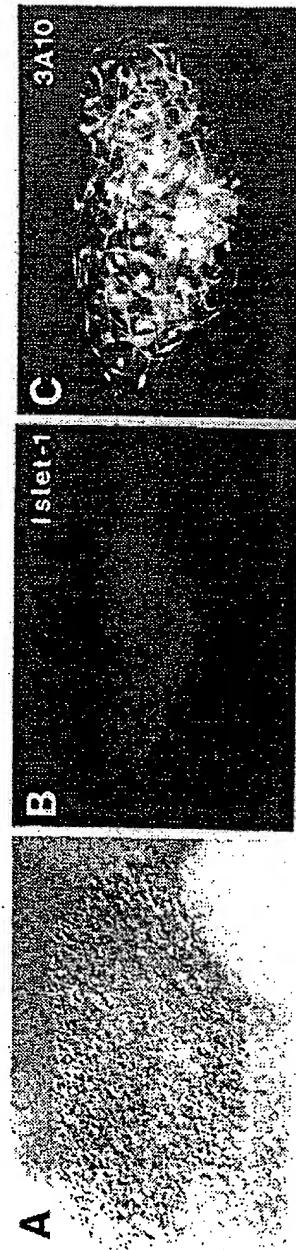


FIGURE 7B

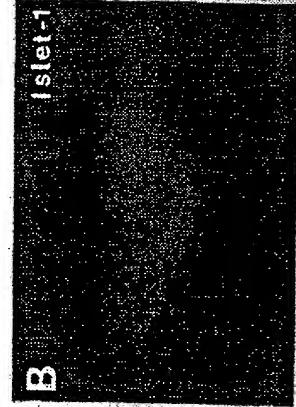


FIGURE 7C

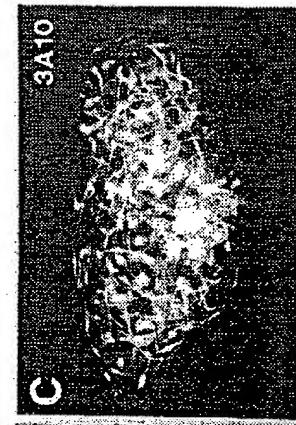


FIGURE 7D

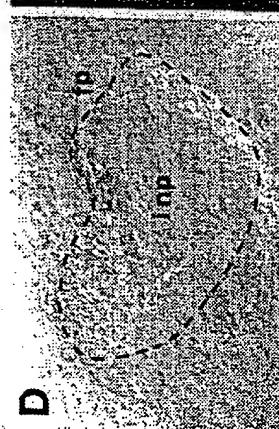


FIGURE 7E

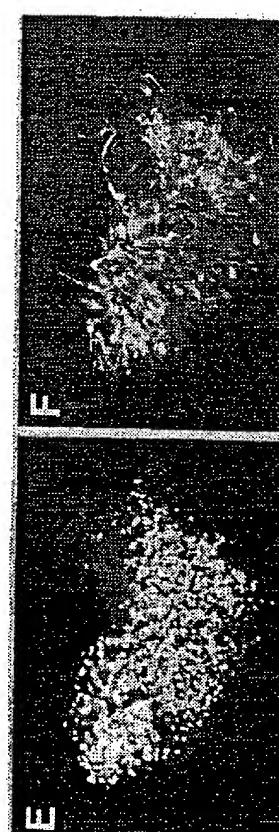


FIGURE 7F



13/16

FIGURE 7G

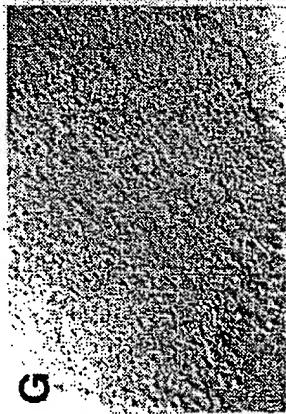


FIGURE 7H

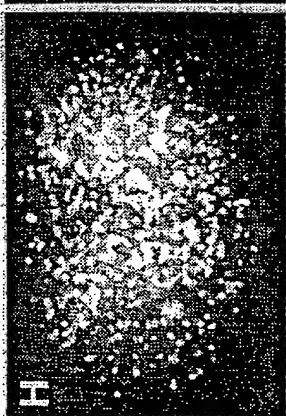


FIGURE 7I

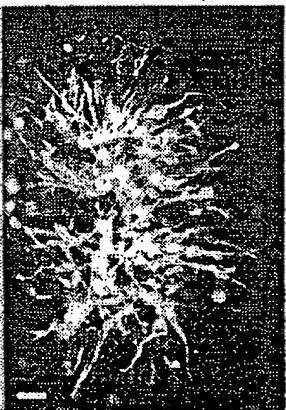


FIGURE 7J

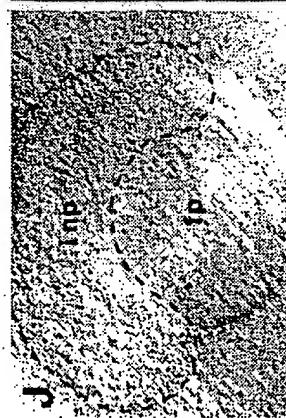


FIGURE 7K

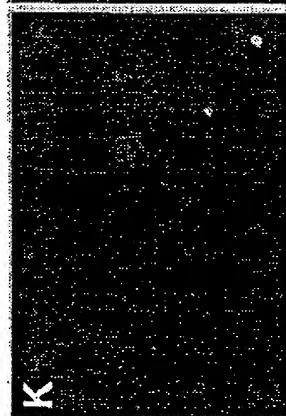


FIGURE 7L



14/16

FIGURE 8A

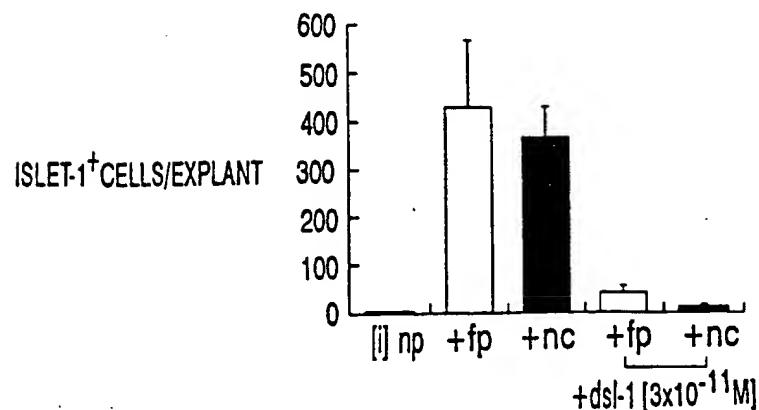


FIGURE 8B

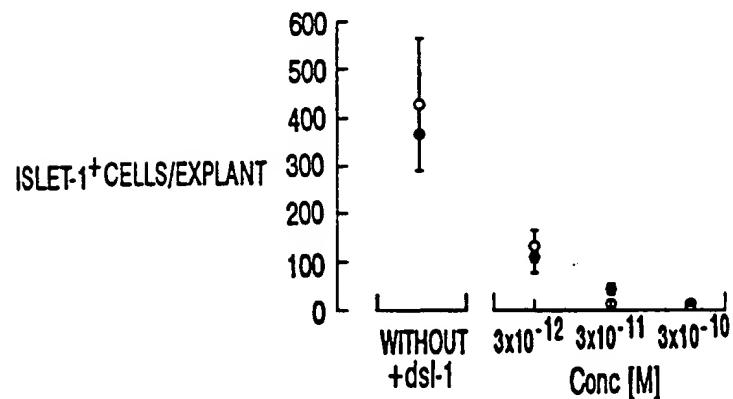
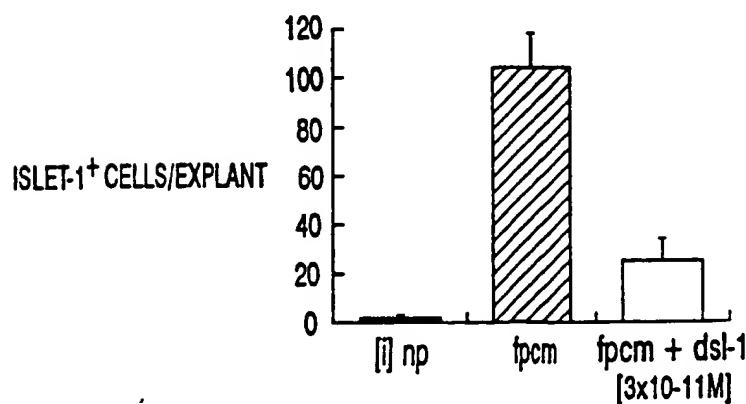


FIGURE 8C

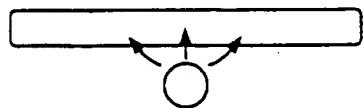


15/16

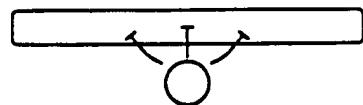
FIGURE 9A

ESTABLISHMENT OF DORSALIN-1 EXPRESSION

i) SIGNALS FROM THE NOTOCHORD SPECIFY THE VENTRAL FATE OF OVERLYING NEURAL PLATE CELLS



ii) SIGNALS FROM THE NOTOCHORD ACT ON OVERLYING NEURAL PLATE CELLS TO PREVENT SUBSEQUENT DSL-1 EXPRESSION



iii) RESTRICTED DORSAL EXPRESSION OF DSL-1 OCCURS AFTER NEURAL TUBE CLOSURE

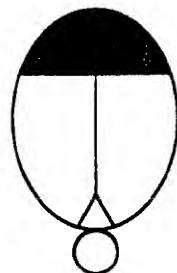
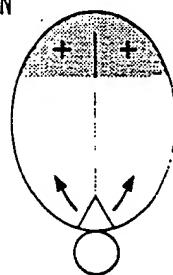


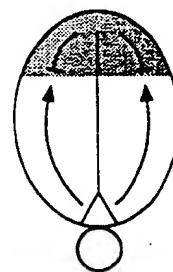
FIGURE 9B

POSSIBLE FUNCTIONS OF DORSALIN-1

i) PROMOTION OF DORSAL CELL TYPE DIFFERENTIATION



ii) LIMITING THE SPREAD OF VENTRAL SIGNALS



iii) DIFFUSION OF DSL-1 CONTROLS CELL PATTERN MORE VENTRALLY

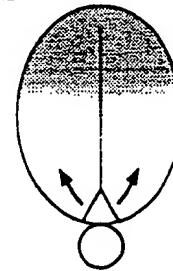


FIGURE 10

| | | | |
|------|-----|---|-----|
| 1 | B29 | HIXFGVLAALSVFNIIACLTGKPLENWKKLPPVMEESDAFFHDPEVEHDTHDFKSFLNMKTDLLRSLNLSRVP SQVK | 80 |
| B29m | ... | ... | ... |
| 81 | B29 | TKEEPPQFTIDLYNRYTADKSSIPASNIVRSFSTEDVVS LISPEEHSFQKHHILLFNISIPRYEEVTRAELRIFISCHKEV | 160 |
| B29m | ... | ... | ... |
| 161 | B29 | GSPSRLEGNNVYDVL.DGDHWNKESTKSLLVSHSIQDCGWEVSSAVKRWVKADKMKTKNKLLEVVIESTKDLGFP | 240 |
| B29m | ... | DVLEDSETWDQATGTTFLVSQDPIRDEGWETLEVSSAVKRWVRADSTTNKLNKLEVTVQSHRES..C | ... |
| 241 | B29 | GRIDITVTHDTKNIPLLIVTSNDRSNGTKETKVE.LREMIVHEQESVLNKLGKNDSSSEEQREEKAI...ARPRQHSSR | 320 |
| B29m | ... | DTLDISVPPGSKNULPFFVFTSNDRSNGTKETRDLILKEMIGHEQETMLVKTAKNAYQGAGESQEEEGLDGYTAVGPLLAR | ... |
| 321 | B29 | SKRSIGA.NHCRRTSLHVNFKEIGWDSWIIAPKDXEAFECKGGCCFP LTDNVTPPTKHAIVQTLVHLQNPKKASKKACCVPT | 400 |
| B29m | ... | RKRSTGASSSHCOKTSLRVNFEDIGWDSWIIAPKEYDAYECKGGCCFP LADDVTPPTKHAIVQTLVHLKFPTKVGKACCVPT | ... |
| 401 | B29 | KIDAIISILYKDDAGVPTLILYEGMKVAECGCR | 433 |
| B29m | ... | KLSPISILYKDDAGVPTLILYEGMKVAECGCR | ... |

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/05743

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :C07K 13/00, 15/28; C12N 15/00; C12P 21/02; A61K 37/02
US CL :530/350, 388.1; 536/23.1; 435/ 69.1, 240.1, 320.1; 514/12

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 530/350, 388.1; 536/23.1; 435/ 69.1, 240.1, 320.1; 514/12

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DIALOG, MEDLINE, WPI

search terms: Dorsalin-1, neural crest cell differentiation, nerve cell regeneration

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category* | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|-----------------------|---|------------------------|
| X, P ----- Y, P | Cell, Volume 73, issued 21 May 1993, Basler et al, "Control of Cell Pattern in the Neural Tube: Regulation of Cell differentiation by <i>dorsalin-1</i> , a Novel TGF β Family Member", pages 687-702, see pages 687-696. | 1-20 ----- 21-40 |
| Y | Cell, Volume 71, issued 30 October 1992, Ferguson et al, " <i>decapentaplegic</i> Acts as a Morphogen to Organize Dorsal-Ventral Pattern in the Drosophila Embryo", pages 451-461, see pages 451 and 452. | 1-40 |
| | | |

 Further documents are listed in the continuation of Box C. See patent family annex.

| | | |
|---|-----|--|
| Special categories of cited documents: | "T" | later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention |
| "A" document defining the general state of the art which is not considered to be of particular relevance | "X" | document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone |
| "E" earlier document published on or after the international filing date | "Y" | document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art |
| "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) | "&" | document member of the same patent family |
| "O" document referring to an oral disclosure, use, exhibition or other means | | |
| "P" document published prior to the international filing date but later than the priority date claimed | | |

Date of the actual completion of the international search

04 AUGUST 1994

Date of mailing of the international search report

15 AUG 1994

Name and mailing address of the ISA/US
Commissioner of Patents and Trademarks
Box PCT
Washington, D.C. 20231
Facsimile No. (703) 305-3230

Authorized officer

Sally Teng

D. Kiyazawa

Telephone No. (703) 308-0196